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Local Correlated Motions in Aqueous Solution of Sodium Chloride*

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Real-space motions of molecules and ions in an aqueous solution of sodium chloride are studied with high-resolution inelastic X-ray scattering. We focus on the correlated dynamics of water molecules and salt ions, instead of the instantaneous structure and the rotational and reorientation dynamics that have been studied using scattering techniques and optical spectroscopy, respectively. The real-space analysis in terms of the Van Hove function illustrates the effect of salt ions to slow down the local correlated dynamics of surrounding water in the hydration shells.

I. INTRODUCTION

Aqueous solutions of salt and electrolyte solutions are ubiquitous and of fundamental importance in many applications and science, including life-sciences. The hydration shells of biomolecules strongly influence their structure and properties [1], whereas the physical properties of electrolyte solutions are key to achieving high performance in batteries. In addition, 97.5 % of water on Earth is salt water [2]. Many studies have been conducted to understand the mechanism of changes in various properties with the addition of salt. For example, the addition of salt to water induces a change in viscosity in an ion-specific manner and shows both increases and decreases upon the addition. A semi-empirical description of this behavior is given by the Jones–Dole equation [3, 4], in which the so-called B -coefficient represents the linear increase and decrease in viscosity with concentration, but its understanding lacks microscopic and molecular picture. Despite significant experimental and theoretical efforts [5–18], a molecular-level understanding of salt water, particularly its transport properties, is elusive and

remains controversial [19]. Conventional neutron/X-ray scattering makes it possible to obtain an instantaneous (snapshot) structure of water molecules in salt solution [5, 13, 20–24], while optical spectroscopy provides us with translational, rotational, and reorientation dynamics of water molecules [6–11, 25–28]. Molecular dynamics (MD) simulation have played a major role regarding visualizing molecular motions in a salt solution to complement the experimental observations [29–31]. However, water models are still far from perfect and sometimes fail to reproduce experimental results even qualitatively [32, 33]. Indeed, the current theory and molecular dynamics simulation cannot predict the values of viscosity even qualitatively nor explain the mechanism of the decrease in the viscosity. Recent spectroscopy studies found the correlation between B -coefficients and various experimental quantities [7, 8] and a coarse-grained model of electrolyte solutions successfully reproduces the basic trend of transport properties [17, 34]; still, its molecular-level understanding of this phenomenon has been elusive.

One of the issues is incomplete understandings of the collective motions of molecules, which are directly relevant to the transport properties of liquid. So far, the difficulty for obtaining the molecular-level understanding lies in identifying the collective molecular motions of water experimentally, in a time-scale relating to the forming/breaking of a hydrogen bond around picoseconds. We have previously identified that the local collective motions of molecules play a vital role in determining the viscosity in liquid. At high temperatures, the time required for an atom losing or gaining one neighbor is a critical quantity that determines the viscosity of liquid [35]. This local configurational excitation time was found to be almost equal to the Maxwell relaxation time, τ_M , for simple metallic liquids [35] and pure water [36, 37] at temperatures above a cross over temperature, T_A , which defines the crossover from the Arrhenius behavior to the

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non-Arrhenius behavior [38]. Here, $\tau_M = \eta/G_\infty$, determines the time scale of viscous phenomena, where η is viscosity and G_∞ is instantaneous shear modulus. The fact that the equality holds for these entirely different properties, one macroscopic (τ_M) and the other microscopic (τ_{LC}), of liquids suggests that this is universal and the process of losing/gaining one neighbor is an elementary excitation in liquids [39].

To shed light on the collective and correlated motions of molecules by experiment, we have employed a new technique: analysis of liquid dynamics using the Van Hove function [40]. The molecular Van Hove function, $G(R, t)$, is defined as

$$G(R, t) = \frac{1}{4\pi\rho NR^2} \sum_{i,j} \delta(R - |\mathbf{r}_i(0) - \mathbf{r}_j(t)|) \quad (1)$$

where ρ is the average number density of molecules, N is the number of molecules in the system, and $\mathbf{r}_i(t)$ is the position of the i -th molecule at time t . The Van Hove function represents the probability for a molecule at the origin $R = 0$ at time $t = 0$ finding another molecule at distance R , at time t , and is regarded as time-dependent pair distribution function (PDF). Thus, the real-space molecular correlated motions can be visualized using the Van Hove function. Previously, it was difficult to obtain Van Hove function experimentally because it requires us to obtain inelastic X-ray/neutron scattering data over wide ranges of energy (E) and momentum transfer (Q) with a high E - and Q -resolution. Recent progress in a high-resolution inelastic X-ray scattering (IXS) using highly-brilliant synchrotron radiation makes it possible to obtain IXS spectra over sufficiently wide ranges of E and Q so that $G(R, t)$ can be calculated via the double Fourier transformation of the IXS spectra over E and Q [41, 42]. This technique has shown the detailed correlated motions of molecules in a picosecond time-scale and its close relationship with macroscopic viscosity in pure water [36, 37] as well as those of simple metallic liquids [43, 44].

In this paper, we employ this technique to study NaCl aqueous solutions with the aim of examining the effect of Na^+ and Cl^- ions on the dynamics of water molecules in real-space. The Van Hove functions are successfully obtained and show the correlated motions of water molecules and ions; the slowing-down of correlated dynamics upon the addition of NaCl is observed. Furthermore, the detailed analysis provides the information about the interaction between water molecule and ions. This research paves the way for elucidating the molecular-level mechanism of water dynamics in the presence of various salts.

II. MATERIALS AND METHODS

A. Materials and Inelastic and elastic X-ray scattering of salt water

Sodium chloride (Fisher Scientific) was used as received. Salt solutions were prepared in approximately 30 cm³ of fresh deionized water, shaken by hand, and were allowed to sit overnight before experiments. The molality of a solution, m , was calculated to be 0.75, 1.5, 2.26, and 3.0 mol/kg.

Inelastic X-ray scattering measurements were carried out at the RIKEN Quantum NanoDynamics Beamline (BL43LXU) [45], SPring-8, in Hyogo, Japan. Salt solution samples were placed between two single crystal diamond windows (thickness: 0.3 mm) and the thickness of sample was 3.0 mm ($m = 0, 0.75, 1.5, 2.26$ mol/kg) or 1.0 mm ($m = 3.0$ mol/kg). The temperature of the sample was kept at 303 K. The energy of incident X-ray was 21.747 keV and the IXS spectra were collected over a wide Q - E range ($0.94 \text{ \AA}^{-1} < Q < 10.1 \text{ \AA}^{-1}$, $-10 \text{ meV} < E < 100 \text{ meV}$ for the samples of $m = 0, 0.75, 1.5$, and 2.26 mol/kg, and $-10 \text{ meV} < E < 70 \text{ meV}$ for the sample of $m = 3.0$ mol/kg). The Q -resolution was $0.4 - 1 \text{ \AA}^{-1}$. To estimate the energy resolution, the IXS spectra were measured for borosilicate glass of 3 mm thick.

To normalize the IXS spectra, the scattering intensity profiles of salt water solution, $I(Q)$, were separately measured at 6ID-D beamline, APS (Argonne National Laboratory, U.S.A.). The data were taken with 100.241 keV X-ray with an amorphous silicon flat panel detector (XRD 1621, Perkin-Elmer, Inc.), and the sample-to-detector distance was 395 mm. Two-dimensional high-energy X-ray diffraction patterns were converted into one-dimensional scattering intensity profiles using *Fit2D* [46]. After completing the conventional correction procedure such as absorption correction and subtracting Compton scattering [47], the profiles of absolute scattering intensity were used for normalizing the IXS data.

B. Calculation of the Van Hove function from IXS data

The measured IXS data, $I_{\text{raw}}(Q, E)$, were reduced to the Van Hove function, $G(R, t)$, by the double Fourier transformation over momentum transfer Q and energy transfer E , following the procedures discussed in the previous studies [36, 37]. In the analysis of pure water, we used a molecular form factor, $f_{\text{H}_2\text{O}}(Q)$, as in the previous studies to obtain the structure factor, $S(Q, E)$, and thereby the molecular Van Hove function, $G(R, t)$, from the scattering intensity $I(Q, E)$: $S(Q, E) = I(Q, E)/\langle f_{\text{H}_2\text{O}}(Q) \rangle^2$. In the analysis of salt water, we divided $I(Q, E)$ by the square of the averaged molecular

form factor, $\langle f(Q) \rangle^2$, defined as follows

$$\langle f(Q) \rangle = c_{\text{H}_2\text{O}} \langle f_{\text{H}_2\text{O}}(Q) \rangle + c_{\text{Na}^+} \langle f_{\text{Na}^+}(Q) \rangle + c_{\text{Cl}^-} \langle f_{\text{Cl}^-}(Q) \rangle \quad (2)$$

where c_i is the atomic concentration of species i whose Q -dependent X-ray molecular/ionic form factor is $f_i(Q)$, and the angular bracket represents the azimuthal averaging. Here we assumed a simple ionic picture that the ions are fully dissolved in water and the form factor of each ion is calculated using a free ion approximation [48]. The molecular form factor of water is taken from the reference [49]. After that, the Van Hove function is calculated by the double Fourier transformation of $S(Q, E)$. As a result, $S(Q, E)$ and $G(R, t)$ represent the total spatiotemporal correlation among water molecules, cations and anions, the detailed description of which is discussed in the following subsection.

The effect of energy resolution is removed in the time-domain, instead of the energy domain as in the conventional IXS studies. The IXS spectra of Borosilicate glass of 3 mm thick was used as the energy resolution function, $M(Q, E)$. We then calculated its Fourier transform over E , $m(Q, t)$ for which Q is at the first peak of scattering function. The decaying behavior of the resolution function was fitted by using the summation of a Gaussian function and an exponential function, as expected from a pseudo-Voigt approximation of the energy resolution function in the energy domain: $m(Q, t) = m_1 \exp(-t^2/\tau_G^2) + m_2 \exp(-t/\tau_E)$. The fitting results showed that the $\tau_G = 0.17$ ps and $\tau_E = 3$ ps with $m_1 = 0.09$ and $m_2 = 0.91$. Assuming $m(Q, t)$ is not highly dependent on Q , the effect of energy resolution function is reasonably removed by dividing the intermediate scattering function, $F(Q, t)$, by $m(Q, t)$. The validity of the assumption will be addressed in a separate study.

C. Partial Van Hove function

In PDF analysis of a multi-component system, several kinds of structure factor and corresponding PDFs have been proposed. Here we introduce a partial dynamic structure factor and the partial Van Hove function following the Faber-Ziman formalism of partial structure factor. The partial Van Hove function between the α and the β , $G_{\alpha\beta}(R, t)$, is defined as

$$G_{\alpha\beta}(R, t) = \frac{V}{4\pi N_\alpha N_\beta R^2} \sum_{i \in \alpha} \sum_{j \in \beta} \delta(R - |r_i(0) - r_j(t)|) \quad (3)$$

where $\{\alpha\}$ represent the atoms (or molecules or ions) of α species. V is the volume and N_α and N_β are the number of molecules of α and β species, respectively. With this definition, $G_{\alpha\beta}(R, t) \rightarrow 1$ with $R \rightarrow \infty$ and the total Van Hove function is given by $G_{\text{tot}}(R, t) = \sum_{\alpha\beta} N_\alpha N_\beta G_{\alpha\beta}(R, t) / \sum_\alpha N_\alpha$. Equivalently, we can define the dynamic structure factor, $S_{\alpha\beta}(Q, E)$, as the

Fourier transform of the partial Van Hove function, $S_{\alpha\beta} = \mathcal{F}(G_{\alpha\beta})$, such that $S_{\alpha\beta}(Q, E) \rightarrow 1$ with $Q \rightarrow \infty$ and

$$\sum_{\alpha, \beta} w_{\alpha\beta}(Q) S_{\alpha\beta}(Q, E) = S(Q, E). \quad (4)$$

Here, the weighting factor, $w_{\alpha\beta}(Q)$, is given by the products of concentration c_i and form factor $f_i(Q)$ of i -th species:

$$w_{\alpha\beta}(Q) = \frac{c_\alpha c_\beta f_\alpha(Q) f_\beta(Q)}{(\sum_\alpha c_\alpha f_\alpha(Q))^2}. \quad (5)$$

Figure 1 shows $w_{\alpha\beta}(Q)$ for the aqueous solution samples used in this study. Because of the Q -dependence of $w_{\alpha\beta}(Q)$, the calculation of the partial Van Hove function from the experimental results is not straightforward. Meanwhile, our measurement at a single incident X-ray energy does not allow us to separate each individual contribution $S_{\alpha\beta}(Q, E)$ from the measured dynamic structure factor. Also, the Q -dependence of $w_{\alpha\beta}(Q)$ is much smaller compared to that of the form factor $f_\alpha(Q)$ as shown in Fig. 1. Therefore, we employ the approximation of weight factors proposed by Warren and introduce

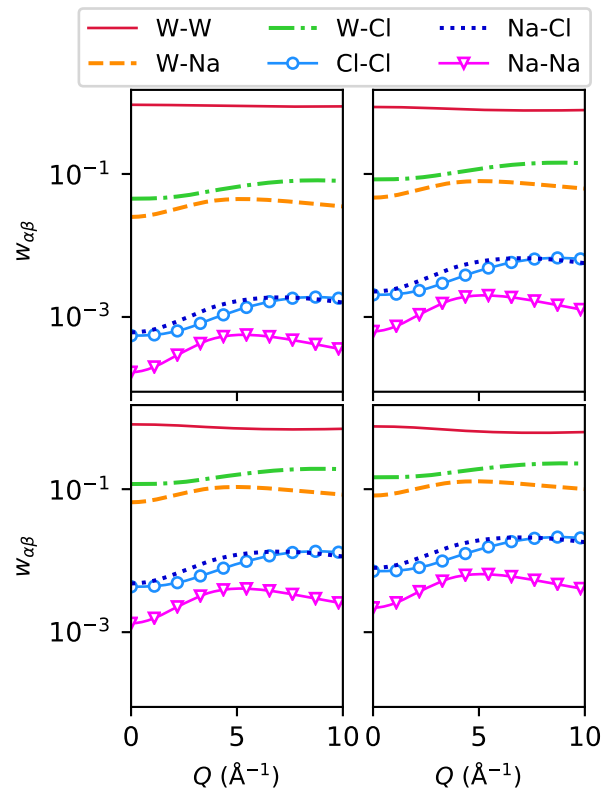


FIG. 1. Comparison of $w_{\alpha\beta}(Q)$: $m =$ (upper left) 0.75, (upper right) 1.5, (lower left) 2.26, and (lower right) 3.0 mol/kg. W stands for water molecule.

the averaged weight factor defined by

$$\overline{w_{\alpha\beta}} = \frac{1}{Q_{\max}} \int_0^{Q_{\max}} w_{\alpha\beta}(Q) dQ. \quad (6)$$

Here, we used $Q_{\max} = 10.0 \text{ \AA}^{-1}$ and the results for Water–Water, Water– Na^+ , and Water– Cl^- are shown in Tab. I. The other correlation is omitted because they are much smaller as clearly seen in Fig. 1. Then, the Van Hove function which is calculated by the double Fourier transformation of $S(Q, E)$ can be expressed as the combination of the pseudo partial Van Hove functions, $G'_{\alpha\beta}(R, t)$, as follows:

$$G(R, t) - 1 = \sum_{\alpha\beta} \overline{w_{\alpha\beta}} (G'_{\alpha\beta}(R, t) - 1). \quad (7)$$

Note that $G'_{\alpha\beta}(R, t)$ is slightly different from the partial Van Hove function, $G_{\alpha\beta}(R, t)$, defined in eq. (3). Even though this is a rough estimation, it is useful enough for estimating the amount of contribution from each correlation to the calculated Van Hove function.

III. RESULTS AND DISCUSSION

A. Van Hove function of salt water

Figure 2 shows the Van Hove functions of pure water and the aqueous solutions of NaCl. As shown in the previous study, the shift in the first and second peak is clearly seen for the pure water; the first neighbor peak position moves with time toward a higher R and the second peak position moves toward a lower R . This is in sharp contrast to the case of metallic liquid, for which the peak positions remain mostly unchanged [36, 50]. The shift indicates that the first and second neighbors are dynamically coupled due to the existence of a hydrogen bond and the small coordination number [36, 37].

Addition of salt induces the changes in the shape of Van Hove function profiles. Regarding the snap-shot structure ($t = 0$), the amplitude of correlation functions decreases as the increase in the salt concentration as is expected from the previous neutron scattering studies [20]. These changes may be interpreted as the result of phase mismatching in the oscillation of the Van Hove function: the position of the individual radial distribution functions for O–O, O– Na^+ , and O– Cl^- are estimated to be around 2.8 \AA , 2.3 \AA , and 3.2 \AA , respectively [21]. In the

current measurement, all of these contribute to the Van Hove function. These values of ionic radii are close to each other but are sufficiently different so that the scattering from each component is canceled out or smeared due to the phase mismatch on the oscillation. An enlarged view at around the first neighbor peak (Fig. 3) indicates the emergence of the correlation between water and Cl^- around the distance which corresponds to the sum of the ionic radii of O^{2-} and Cl^- .

At a longer time-scale ($t > 0.5$ ps), the addition of salt induces a qualitative change at the first and the second neighbor peaks. The addition of salt makes the decay time of the first neighbor peak longer as is discussed in detail in the next section, whereas the amplitude of the second neighbor peak is drastically decreased by salt. Therefore, the first neighbor correlation is dominant in the NaCl aqueous solution at $t < 1.0$ ps. This is in stark contrast to the case of pure water, where second neighbor peak at around $R = 4.2 \text{ \AA}$ is dominant after the first neighbor peak at around $R = 2.8 \text{ \AA}$ fully decays at around $t = 1\text{--}1.5$ ps [37].

Because of the intrinsic broad width of correlation profiles, it is difficult to separate the profiles into those of each individual oxygen–ion correlations a priori. Therefore, we first discuss the overall correlated motions between neighboring water molecules and salt ions. Then, we discuss the contribution of oxygen–ion correlations by focusing on the variation in the Van Hove function induced by salt ions.

B. Temporal evolution of the first neighbor peak

The decaying behavior of the first neighbor peak, $R = R_1$, is directly related to the correlated motions between neighboring molecules, thereby largely determining the transport phenomena such as viscosity as shown in the previous studies [37]. We find that the addition of salt induces the changes in the decaying behavior of the peak area, $A_1(t)$, at the first neighbor peak as shown in Fig. 4. Here, the peak area is defined as $A_1(t) = \int_{R_1'}^{R_1''} (G(R, t) - 1) dR$, where R_1' and R_1'' are defined as shown in Fig. 3. To estimate the uncertainties, the upper bound R_1'' was changed within the gray-shaded region in Fig. 3.

Pure water shows a two-step relaxation behavior as observed in the previous study [37]. The two-step decay is also observed for the samples of aqueous solution of NaCl, but with the addition of salt, the amplitude of the first step decay is decreased (see the inset of Fig. 4), while that of the second-step decay is increased except for the highest concentration sample ($m = 3.0$ mol/kg), which shows a decay similar to that of the 2.26 mol/kg sample. The choice of R_1'' is somewhat arbitrary, but the uncertainties that were estimated by changing R_1'' hardly influence the following discussion.

This two-step decaying behavior is fitted by using the summation of two compressed exponential functions,

TABLE I. List of the averaged weight factor.

Molality (mol/kg)	0.75	1.5	2.26	3.0
Water–Water	0.894	0.804	0.728	0.663
Water–Na	0.038	0.068	0.092	0.111
Water–Cl	0.065	0.116	0.157	0.189

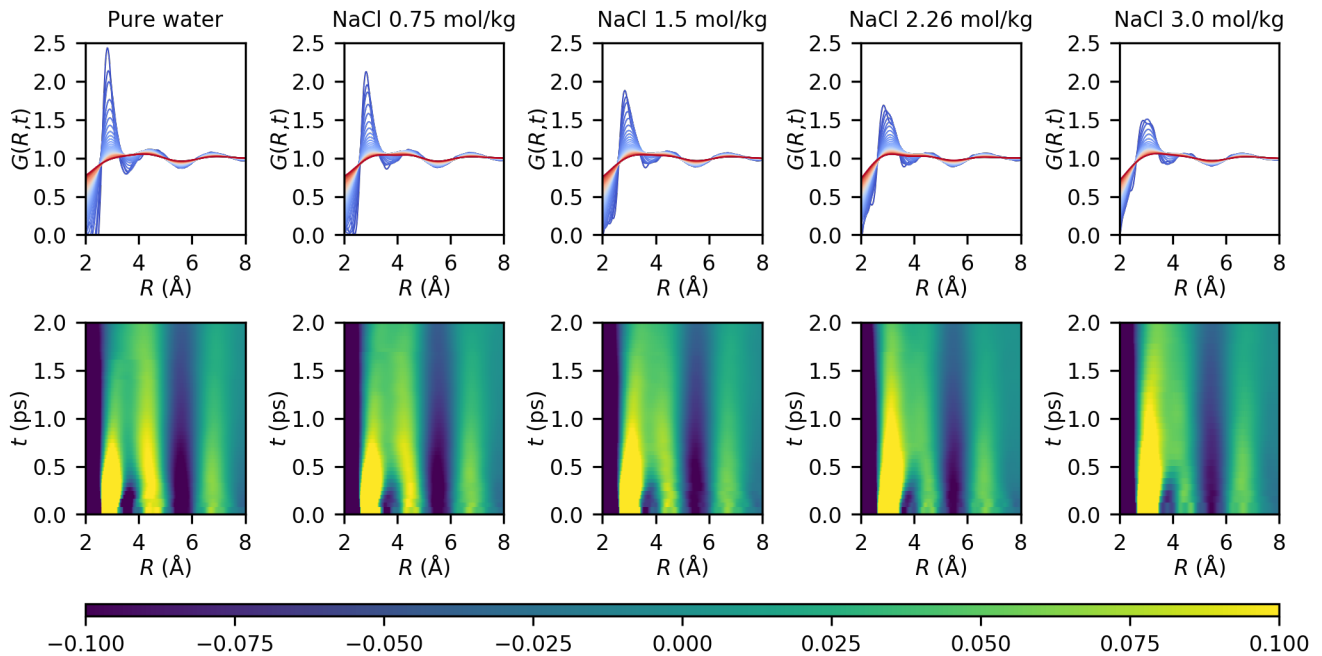


FIG. 2. Van Hove function of NaCl aqueous solution. (Top) One-dimensional slice of the Van Hove functions, $G(R, t)$, at $0 < t < 2$ ps. (Middle) Intensity map of $G(R, t) - 1$, the color scale of which is shown in the bottom. (Left to right) $m = 0$ (pure water), 0.75, 1.5, 2.26, and 3.0 mol/kg.

$A_1(t) = a_1 \exp(-(t/\tau_1)^{p_1}) + a_2 \exp(-(t/\tau_2)^{p_2})$ with $\tau_1 < \tau_2$. The result of fitting and $A_1(0)$ are shown in Fig. 5. The addition of salt decreases a_1 and increases a_2 , while $A_1(0)$ is decreased. The decrease in $A_1(0)$ indicates a reduction in the apparent coordination number at R_1 . The exponent of the second step decay, p_2 , for the pure water sample is similar to those observed in the previous study [37] and is much higher than that for the aqueous salt solutions.

It has been assumed that the salt influences the dynamics of the first hydration layer, or some extended region beyond the first hydration layer [10, 11]. It is then natural to assume that the correlated dynamics of water molecules that are not close to the ions is identical to that in pure water, while the presence of ions may influence the motions of surrounding water molecules. The amount of volume in which the ions affects the dynamics of water will be proportional to the molality up to m_c at which the hydration layers around ions start to overlap. With this assumption, the decaying behavior of 0.75 and 1.5 mol/kg is expressed as the linear combination of the decaying behaviors of pure water and NaCl solutions in a higher concentration:

$$A_1(t, m) = \frac{2.26 - \frac{m}{\text{mol/kg}}}{2.26} A_1(t, m = 0) + \frac{\frac{m}{\text{mol/kg}}}{2.26} A_1(t, m = 2.26 \text{ mol/kg}) \quad (8)$$

Here we use $A_1(t, m = 2.26 \text{ mol/kg})$ instead of $A_1(t, m = 3.0 \text{ mol/kg})$, simply because apparently the region that

is influenced by the salt is saturated at concentrations higher than 2.26 mol/kg. The result of the linear combination for $m = 0.75$ (dashed line) and 1.5 mol/kg (solid line) is shown in Fig. 4; their agreement with the $A_1(t, m = 0)$ and $A_1(t, m = 1.5 \text{ mol/kg})$ is good enough to conclude that their decay behavior can be expressed as the linear combination of that of the bulk water and the hydration layers. The fact that all the decay curves intersect at $t = 0.28$ ps supports that the spectra are decomposed into the linear combination.

C. Correlation functions between water and ions

The Van Hove function is made of the contributions from the water–water correlations and the water–ion correlations because the ion–ion contribution is negligible at these salt concentrations. The discussion above promotes us to assume that these contributions are independent of the salt concentration, at least in the zero-th order. Then the Van Hove function, $G(R, t)$, may be expressed as the linear combination of the Van Hove function of pure water, $G_{\text{pure}}(R, t)$, and the pseudo partial Van Hove functions for water and ions as follows:

$$G(R, t) - 1 \approx \overline{w_{\text{w-w}}}(G_{\text{pure}}(R, t) - 1) + \overline{w_{\text{w-Cl}}}(G'_{\text{w-Cl}}(R, t) - 1) + \overline{w_{\text{w-Na}}}(G'_{\text{w-Na}}(R, t) - 1) \quad (9)$$

Here, the weighting factors, $\overline{w_{\alpha\beta}}$, are given in Tab. 1 and w stands for a water molecule. Because $\overline{w_{\text{w-Na}}}/\overline{w_{\text{w-Cl}}} \sim$

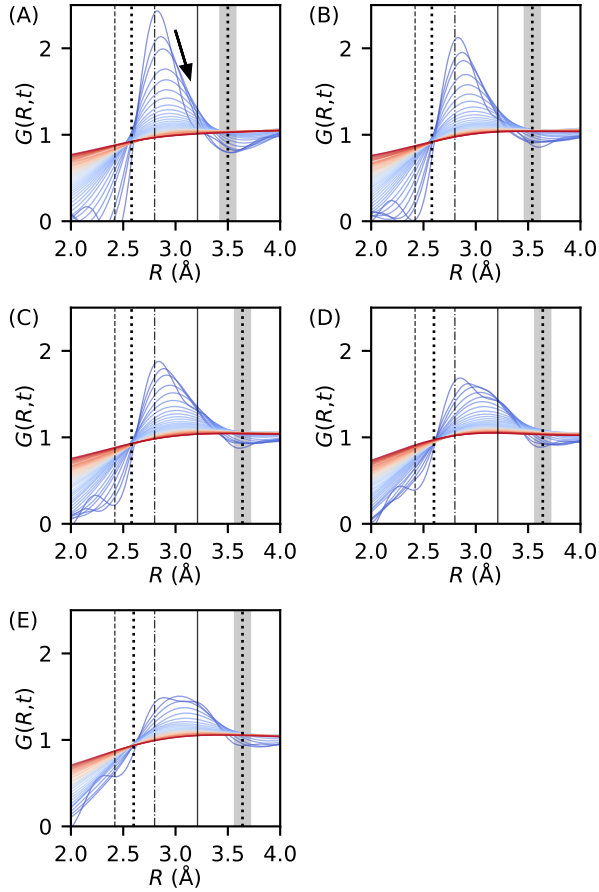


FIG. 3. Enlarged view of Van Hove functions at around the first neighbor correlation peak, $R \sim 2.9\text{\AA}$: (A) pure water, (B) $m = 0.75$ mol/kg, (C) 1.5 mol/kg, (D) 2.26 mol/kg, and (E) 3.0 mol/kg. The solid lines at $R = 3.21\text{\AA}$ show the $R_{\text{O}2^-} + R_{\text{Cl}^-}$. The dashed line at $R = 2.42\text{\AA}$ shows the $R_{\text{O}2^-} + R_{\text{Na}^+}$. The dash-dotted line at $R = 2.8\text{\AA}$ shows the $R_{\text{O}2^-} + R_{\text{O}2^-}$. The range between the dotted lines (R'_1 and R''_1) was used to calculate the area, $A(t)$, of the first neighbor. The upper limit of this range is changed within the gray-shaded area to estimate the uncertainties.

0.58, we may focus on the water-Cl correlation and express the temporal correlations between water and ions by the following:

$$G'(R, t) = \frac{G_m(R, t) - 1 - \overline{w_{w-w}}(G_{\text{pure}}(R, t) - 1)}{\overline{w_{w-Cl}}} \\ \approx (G'_{w-Cl}(R, t) - 1) + \frac{\overline{w_{w-Na}}}{\overline{w_{w-Cl}}}(G'_{w-Na}(R, t) - 1) \quad (10)$$

The time-evolution and its intensity map for each molality are shown in Fig. 6. At a short time ($t < 0.3$ ps), positive correlation peaks are observed at a distance corresponding to $R = R_{\text{O}2^-} + R_{\text{Na}^+}$ ($= 2.42$ Å) and $R = R_{\text{O}2^-} + R_{\text{Cl}^-}$ ($= 3.21$ Å), while a negative correlation peak is observed at $R = R_{\text{O}2^-} + R_{\text{O}2^-}$ ($= 2.8$ Å). The positive correlations represent the correlation between a water molecule and a cation or an anion, respectively, while the negative correlation underscores that there is anti-

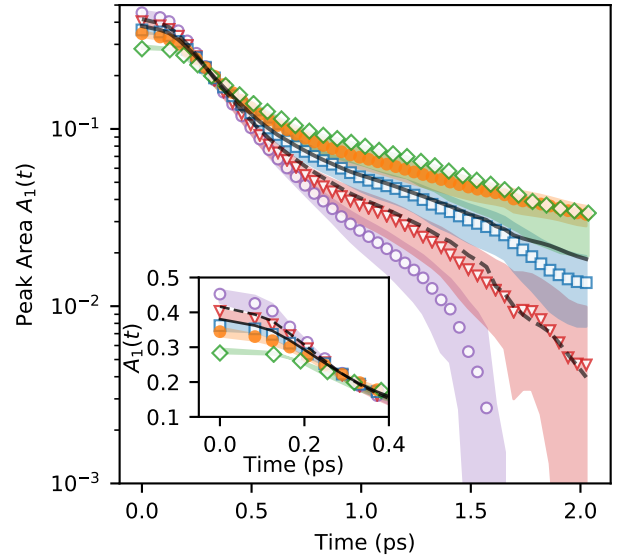


FIG. 4. Temporal changes in the area of first neighbor peak, $A_1(t)$ and the enlarged view (inset): (open circles) pure water, (triangles) $m = 0.75$ mol/kg, (squares) 1.5 mol/kg, (closed circles) 2.26 mol/kg, and (diamonds) 3.0 mol/kg. The shaded areas represent uncertainties of each dataset. The solid and dashed lines represent the linear combination of time-evolution for $m = 0$ and 2.26 mol/kg (see the main text).

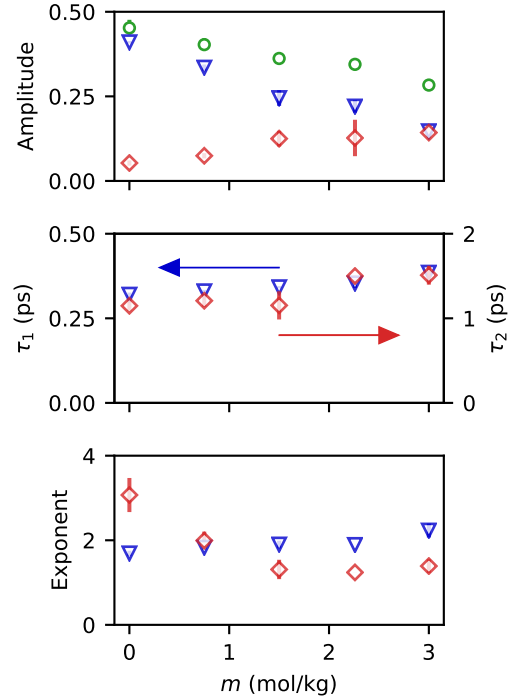


FIG. 5. Fitting Results of the first neighbor decay. (Triangles) a_1 , τ_1 , p_1 , (diamonds) a_2 , τ_2 , p_2 , and (circles) $A_1(0)$. The error bar is estimated by the standard deviation of the fitting.

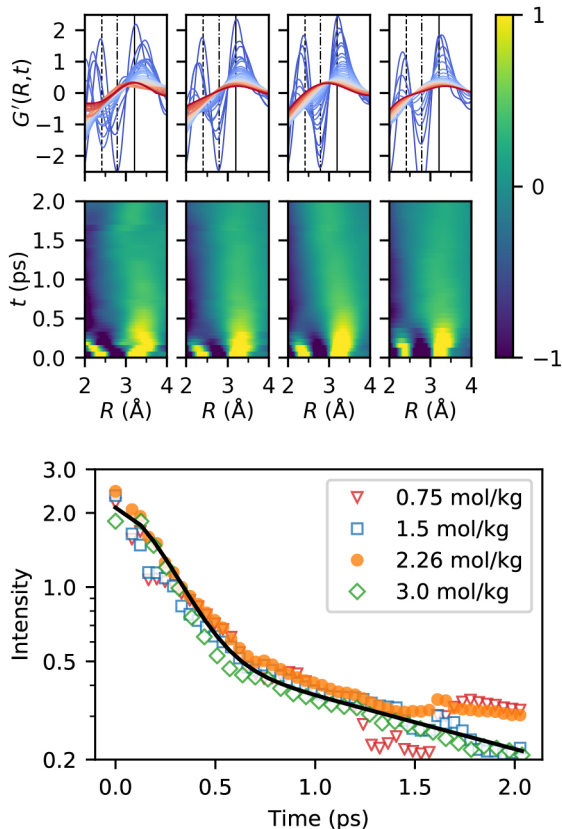


FIG. 6. One-dimensional profiles of $G'(R, t)$ at $0 < t < 2$ ps and (middle) their intensity maps. Their color scale is shown in the right. The molality of sample is 0.75, 1.5, 2.26, 3.0 mol/kg from the left to the right. The solid lines, dashed lines, and the dash-dotted lines in the top figures represent $R = 3.21 \text{ \AA}$ ($R_{\text{O}_2^-} + R_{\text{Cl}^-}$), $R = 2.42 \text{ \AA}$ ($R_{\text{O}_2^-} + R_{\text{Na}^+}$), and $R = 2.8 \text{ \AA}$ ($R_{\text{O}_2^-} + R_{\text{O}_2^-}$), respectively. (Bottom) Time-evolution of peak height at around $R = 3.21 \text{ \AA}$. The solid line shows the result of fitting using two (compressed) exponential functions (see main text).

correlation between water and ions. The peak values of the negative correlation at $R = R_{\text{O}_2^-} + R_{\text{O}_2^-}$ is strongly negative, around -2.5 at $t = 0$, indicating that the correlation between water molecules are perturbed in the hydration shell. It is possible that the local water–water coordination is reduced in the hydration shell, thereby leading to the significantly negative peak values. This interpretation is consistent with the result of neutron diffraction study [13] which concluded that the first peak height in the pair distribution function for water–water correlations is reduced by the addition of NaCl.

Figures in Fig. 6 (top and middle) are similar to each other, underscoring our assumption that the decaying behaviors of water– Na^+ and water– Cl^- correlations are independent of salt concentration, although quantitative discussion is limited due to a contamination of termination errors in the Van Hove functions at a short time, particularly at $t < 0.2$ ps, and the approxima-

tion used for the calculation. The decaying behavior of peak height at around $R = R_{\text{O}_2^-} + R_{\text{Cl}^-}$ (water– Cl^- correlations) is approximated by a two-step relaxation, $I(t) = 1.5 \exp(-(t/0.32 \text{ ps})^{1.7}) + 0.6 \exp(-t/2.0 \text{ ps})$, as shown by the solid line in Fig. 7 (bottom). The first term is almost identical to that of pure water ($\tau_1 = 0.32$ ps) except for its amplitude, indicating that the decaying at a short time scale ($t < 0.3$ ps) corresponds to a ballistic, caged-motion, as was observed for the pure water. The second term represents the presence of a slow-decay in the correlation between water and Cl^- , which dominates the decaying behavior at the first neighbor in Fig. 4. The correlated motions between a water molecule and an Cl^- ion are slower than that of water–water correlations with a decaying time ~ 2 ps, and it is most likely that this correlation is the principal mechanism of the slowing-down shown in Fig. 4. A thorough research using other types of ions would show us how the water–ion correlation depends on the nature of ions.

D. Correlations at a long distance

In contrast to the first and second neighbor peaks, the third neighbor peak at around $R = 6.8 \text{ \AA}$ is hardly dependent on the addition of salt. Figure 7 shows the temporal evolution of the area, $A_3(t)$, at the third peak, which is normalized by $A_3(0)$. The normalized values collapse into a single decaying curve even though $A_3(0)$ gently decreases with the addition of salt. This indicates that the dynamics of the NaCl aqueous solution is approximately the same at long distances regardless of the molality, which has been discussed in terms of snapshot structure of aqueous salt solution [34], and qualitatively agree with the concept of “sticky ions” [51].

The values of normalized amplitude remain almost constant for $t < 0.3$ ps, indicating that the first-step decay at the first neighbor in the averaged Van Hove functions is absent for the third peak. As was discussed elsewhere [50] the higher order peaks of the PDF and the Van Hove function do not represent a single atom–atom distance, but the density correlations among the aggregate of many atoms. Therefore, the decay time of the higher order peaks does not correspond to the microscopic time-scale. Indeed, the dynamics at the third neighbor peak is barely dependent on the salt concentration.

IV. CONCLUDING REMARKS

Usually the dynamics of liquid is discussed in terms of the dynamic structure function, $S(Q, E)$, or the intermediate scattering function, $F(Q, t)$. In particular, the decay time of the first peak ($Q = Q_1$) of $F(Q, t)$ is called the α -relaxation time and has been widely used to discuss the dynamics of liquid. However, the first peak of $F(Q, t)$ includes all the contributions from the long-

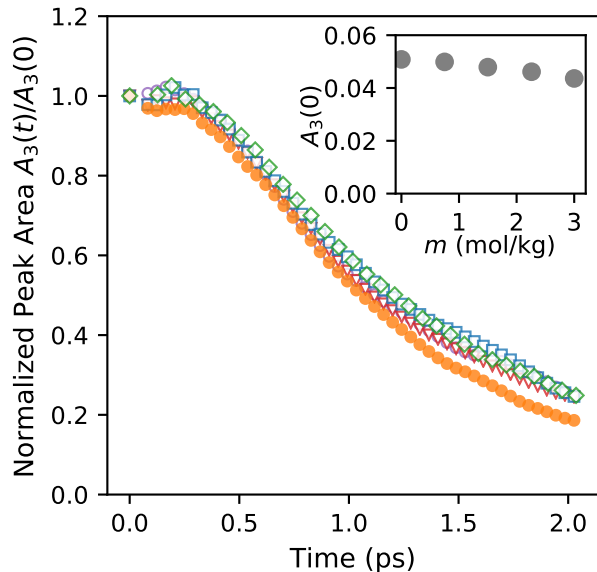


FIG. 7. Normalized temporal changes in the area of third peak, $A_3(t)/A_3(0)$, around $R = 6.8$ Å. (open circles) pure water, (triangles) $m = 0.75$ mol/kg, (squares) 1.5 mol/kg, (closed circles) 2.26 mol/kg, and (diamonds) 3.0 mol/kg. (Inset) Dependence of $A_3(0)$ on the molality.

range correlations, and is dominated by the geometrical factor in many cases rather than the microscopic dynamics [50]. This point underscores the importance of discussing the liquid dynamics in terms of the Van Hove function $G(R, t)$. The effect of salt on the dynamics, within the hydration layers in particular, needs to be examined using the correlation function in real-space, not

in reciprocal-space.

We showed that a real-space visualization of correlated molecular/ionic dynamics in picosecond time-scale and sub-nm scale using the Van Hove function can be applied to aqueous solutions of sodium chloride to determine the effect of salt addition to the local correlated dynamics of water. The results depict how salt ions alter the local correlated molecular motions through the interaction between water and salt ions. The effect is local, not long-range, and is limited to within the hydration cells, which is comparable to what is observed for the reorientation dynamics by optical spectroscopy. Because of the importance of salt water to life and other activities elucidating the effect of salt addition to water at the microscopic level is crucial in advancing the life sciences. The present result represents a major step forward to understanding the transport properties, such as viscosity and conductivity, of complex aqueous solutions based on the molecular-level correlated dynamics.

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