

This is the accepted manuscript made available via CHORUS. The article has been published as:

Thermodynamically consistent treatment of the growth of a biopolymer in the presence of a smooth obstacle interaction potential

F. Motahari and A. E. Carlsson

Phys. Rev. E **100**, 042409 — Published 17 October 2019

DOI: [10.1103/PhysRevE.100.042409](https://doi.org/10.1103/PhysRevE.100.042409)

Thermodynamically consistent treatment of the growth of a biopolymer in the presence of a smooth obstacle interaction potential

F. Motahari and A. E. Carlsson*

*Department of Physics and Center for Engineering Mechanobiology,
Washington University, St. Louis, Missouri 63130, USA*

(Dated: September 27, 2019)

We investigate the effect of filament-obstacle interactions on the force-velocity relation of growing biopolymers, via calculations explicitly treating obstacle diffusion and stochastic addition and subtraction of subunits. We first show that the instantaneous subunit on- and off-rates satisfy a rigorous thermodynamic relationship determined by the filament-obstacle interaction potential, which has been violated by several calculations in the literature. The instantaneous rates depend not only on the average force on the obstacle, but also on the shape of the potential on the nanometer length scale. Basing obstacle-induced reduction of the on-rate entirely on the force, as previous work has often done, is thermodynamically inconsistent and can overestimate the stall force, sometimes by more than a factor of two. We perform simulations and analytic calculations of the force-velocity relation satisfying the thermodynamic relationship. The force-velocity relation can deviate strongly from the Brownian-Ratchet predictions. For shallow potential wells of depth $\sim 5k_B T$, which might correspond to transient filament-membrane attachments, the velocity drops more rapidly than predicted by the Brownian-Ratchet model, in some cases by as much as a factor of 50 at an opposing force of only 1 pN. On the other hand, the zero-force velocity is much less affected than would be expected from naive use of the Boltzmann factor. Furthermore, the growth velocity has a surprisingly strong dependence on the obstacle diffusion coefficient even when the dimensionless diffusion coefficient is large. For deep potential wells, as might result from strong filament-membrane links, both the on- and off-rates are reduced significantly, slowing polymerization. Such potentials can sustain pulling forces while polymerizing, but only if the attractive well is relatively flat over a region comparable to or greater than the monomer size. For double-well potentials, which have such a flat region, the slowing of polymerization by external pushing force is almost linear up to the stall force in some parameter ranges.

1. INTRODUCTION

Polymerization of biopolymers such as actin filaments provides force to drive both protrusion and invagination of the cell membrane. This process is modulated by the interaction between the filament tip and the membrane, which varies considerably between different cellular phenomena. In lamellipodia, filaments are likely weakly bound to the membrane by their interaction with, for example, WASP-family actin-nucleating proteins [1]. On the other hand, in filopodia actin filaments are nucleated by formins, which can bind actin filaments strongly [2]. Furthermore, in processes that require strong pulling forces to bend the membrane, such as endocytosis in yeast, some actin filaments must be strongly bound to the endocytic site. In this process, the binding is believed to result from the protein Sla2, which has both actin-binding domain and a domain that links it to the membrane. [3]. Recent super-resolution experiments have shown that WASP (Las17 in yeast) forms a ring around a Sla2 dot [4], and pulling forces are probably concentrated in the dot [5, 6]. Similarly, microtubule interactions with the cell membrane are mediated by a range of proteins, which may lead to a variety of effective interaction potentials [7, 8]. In all of these cases, it is important to understand how the interaction between a growing filament and the membrane plus associated proteins affects the polymerization rate. The filament-membrane interaction may be optimized for different criteria, such as polymerization velocity where migration speed is crucial, or stability where strong pulling forces are required.

Most previous calculations of force generation by polymerization have used hard wall repulsive potentials acting between the filament tip and the obstacle. The classic Brownian Ratchet (BR) model [9] used such a potential to treat polymerization in the presence of a diffusing obstacle, assuming that the monomer on-rate increases suddenly from zero to its free-filament value at a certain distance from the membrane. Analysis of this model showed that in the limit of fast obstacle diffusion, the hard-wall potential gives rise to a growth velocity that decays exponentially

*Electronic address: aec@wustl.edu

with opposing force:

$$v_{growth} = \delta_{\parallel} [k_{on}^0 \exp(-F\delta_{\parallel}/k_B T) - k_{off}^0], \quad (1)$$

where

$$\delta_{\parallel} = \delta \cos \theta \quad (2)$$

is the step size per added subunit projected on the direction of obstacle motion, θ is the angle of filament growth relative to the direction of motion, δ is the step size and $k_{on,off}^0$ are the free-filament on- and off-rates.

Calculations based on this analysis can qualitatively explain the mechanism behind the generation of pushing forces by actin polymerization in several types of cellular protrusions, such as filopodial and lamellipodial protrusions [10–19]. The basic features of intracellular motion of pathogens such as *Listeria*, which is driven by actin polymerization, can also be explained by BR-type models.

However, the true interaction is continuously varying and the force may have both attractive and repulsive components. It is not known how these variations affect the force-velocity relation. For example, a potential with a deep well might trap the growing end of the filament near the obstacle, and thus slow polymerization. On the other hand, a smoother potential might speed polymerization. A small number of calculations have treated such smoothly varying potentials. Calculations in 2D using an explicitly moving obstacle with explicitly diffusing monomers to treat the growth rate of a single actin filament, interacting with an obstacle via a steeply increasing force field, found that the velocity decays more rapidly than the BR prediction [20]. This effect was attributed to a diffusion barrier in which monomers had to traverse a tunnel-like region to reach their binding site at the end of the filament. Such effects are expected to be smaller in 3D. Calculations using a range of force fields to treat 3D polymerization [21, 22], including filament bending, found large acceleration of growth by a soft obstacle when the obstacle diffusion coefficient was small. Refs. [23] and [24] considered actin filaments strongly bound to the obstacle, but assumed they did not grow. Studies of filament growth while clamped to a motile obstacle via a deep potential energy well suggested that the filament-obstacle attachment is the controlling factor for the elongation rate [25, 26]. A later model considered filaments attached to an obstacle with a double-well potential [27], explicitly treating diffusive motion of the obstacle. It was found that a filament can push the obstacle and grow with a speed of about half of the free filament speed and thus progressively polymerize, if the potential is sufficiently deep.

Among these preceding studies, there is no systematic exploration of a broad range of possible force fields, to establish how the force field influences the force-velocity relation. In fact, the methodology for performing calculations with smoothly varying filament-obstacle interactions is not well established. One must choose spatial dependences for both the interaction energy between the obstacle and the filament tip, and the polymerization rate parameters. In previous work these dependences have often been chosen independently.

Here we first establish a thermodynamic relationship between the spatial dependence of the polymerization-rate parameters and that of the interaction energy, simplifying the construction of appropriate force-generation models. Several preceding studies are inconsistent with this relationship. The relationship applies to a filament growing against a diffusing obstacle whose motion is treated explicitly, with a smoothly varying filament-obstacle interaction. Calculations not implementing this relationship can overestimate the stall force. We then perform a systematic set of simulations for a broad range of possible filament-obstacle interaction potentials, treating polymerization and depolymerization as well as obstacle motion stochastically. We find that monotonically decaying repulsive potentials lead to force-velocity relations very similar to the BR prediction, as expected from general thermodynamic principles. But in other cases the results can differ strongly from the BR prediction. Weak attractive potentials lead to much faster decay than is predicted by the BR model, and reduce the zero-force velocity. Deep and narrow potentials lead to slow polymerization at all force values. Calculations of the maximum pulling force and growth velocity as functions of well depth show that attached filaments stop growing if the potential has a single narrow well of nanometer-scale width that is deep enough to sustain pulling forces greater than about 1pN. Wider potentials, such as the double-well potential that we consider, can have rapid polymerization at zero force, and have a force-velocity relation that decays almost linearly for pushing forces up to the stall force.

Although the model is highly simplified, multiscale calculations such as those of Ref. [10, 12, 19, 28–32] have demonstrated the utility of simple, but approximate results for the force-velocity relation in calculating the properties of cells and processes inside cells. The general understanding gained from the present studies will enhance the physical relevance of such multiscale calculations.

2. MODEL

2.1. Filament-Obstacle Interaction

The model (Figure 1) treats the stochastic polymerization of a biopolymer, exerting force on an explicitly moving, flat, penetrable obstacle. The filament grows at an angle θ relative to the obstacle direction of motion. We envisage the base of the filament as being rigidly anchored. Actin filaments, for example, could be anchored in a crosslinked actin meshwork. The growing end of the filament is assumed to interact with a membrane-binding protein that attaches it to the membrane. Examples of such proteins include i) Hip1R (mammals) and Sla2 (yeast), which link actin filaments strongly to the membrane as part of the pulling-force generation process during endocytosis, and ii) WASP, which tethers filaments weakly to the membrane during the growth of dendritic actin networks [33].

The details of the interaction potentials between the actin-filament tips and the membrane proteins are not known. Therefore, we treat a range of filament-obstacle interactions given by smooth potential functions having the form

$$U(r) = Ae^{-\kappa_1 r} - Be^{-\kappa_2 r} \quad (3)$$

or

$$U(r) = Ae^{-\kappa_1 r} - Ce^{-[\kappa_3(r-r_1)]^2} - De^{-[\kappa_4(r-r_2)]^2} \quad (4)$$

Here, A , B , C , D , κ_1 , κ_2 , κ_3 , κ_4 , r_1 , and r_2 are constants, and r is the variable gap between the tip of the filament and the obstacle. Filament-obstacle binding can be naturally included in the force field via the B term in Eq. 3; we denote such a potential a “simple well”. A non-zero C in Eq. 4 also adds a Gaussian “spike” to the potential at $r = r_1$ which can be either attractive or repulsive. Choosing both C and D to be positive in Eq. 4 generates a double-well potential, as shown in Figure 2(b). Here we have chosen $r_1 = 2.45$ nm and $r_2 = 4.85$ for illustration; we find that the force-velocity relation is not sensitive to their precise values as long as $r_2 - r_1$ is close to filament length added per new subunit. The depth of the wells in the potentials represents the energy of binding filaments to membrane-bound proteins. The corresponding forces that the filament exerts on the obstacle are

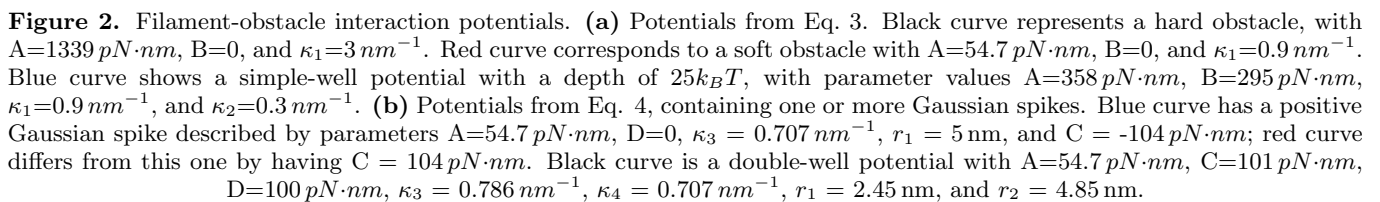
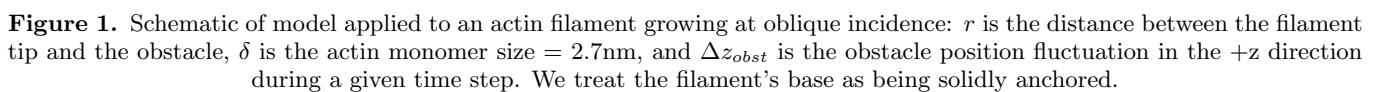
$$F(r) = -\frac{dU}{dr} = A\kappa_1 e^{-\kappa_1 r} - B\kappa_2 e^{-\kappa_2 r} \quad (5)$$

and

$$F(r) = A\kappa_1 e^{-\kappa_1 r} - 2C\kappa_3^2(r - r_1)e^{-[\kappa_3(r-r_1)]^2} - 2D\kappa_4^2(r - r_2)e^{-[\kappa_4(r-r_2)]^2}, \quad (6)$$

respectively. We will denote a filament having only repulsive potential terms a “pusher”. A filament with an interaction potential containing a well or a spike is denoted a “puller” (although it can also exert a pushing force).

We emphasize that the forces calculated here are instantaneous forces rather than being time-averaged. In the case of a hard wall, a time-averaged force between the filament tip and the obstacle can be calculated analytically without performing simulations [34]. However, the calculations that we perform here require knowledge of the force at each time step because, as we shall see below, even nanometer-scale motions of the filament tip relative to the obstacle can cause large changes in the rate constants. Therefore, we perform fully stochastic simulations with fine time resolution. **The position of the filament tip relative to the obstacle is determined by a balance between forces from the filament-obstacle interaction and filament bending (see Sec. 2.3). This force balance will hold over time scales long enough that the random thermal motion averages to zero.**



The obstacle position is stepped forward in time according to biased Brownian motion driven by $F(r)$ and thermal fluctuations. We use a “filament-centric” approach in our simulations, in which the filament base is assumed to be stationary and the obstacle moves. Cases where the filament base is moving can be handled by a simple coordinate transformation, in which obstacle motion is the inverse of the base motion and the diffusion coefficient of the obstacle is replaced by that of the entity to which the filament is anchored. At each time step, the diffusive motion of the

obstacle is calculated by the discrete form of the Langevin equation (Ref. [35], Chap. 3):

$$\Delta z_{obst} = \alpha \sqrt{24\Delta t} \sqrt{D_{obst}} + \frac{D_{obst}}{k_B T} \Delta t [F(r) + F_{ext}] \quad (7)$$

where D_{obst} is the obstacle diffusion constant, F_{ext} is the external force applied on the obstacle, Δt is the time step, and α is a random number uniformly distributed between $-\frac{1}{2}$ and $\frac{1}{2}$, so that $\langle \alpha^2 \rangle = \frac{1}{12}$. Consecutive time steps are uncorrelated.

2.3. Filament-Tip Fluctuations

We show results here including filament-tip fluctuations, and present results in the Appendix excluding filament-tip fluctuations. We treat oblique incidence because the fluctuations are most pronounced in this case, and it is typical for lamellipodia. It is necessary to specify a baseline incidence angle because this determines the projection of the filament-tip fluctuations on the direction parallel to obstacle motion. We use a baseline angle of $\theta = 45^\circ$ as in Figure 1. The model describes fluctuations relative to this baseline angle, whose magnitude is determined by the persistence length of actin. The filament-tip fluctuations are modeled by a variable z_{tip} describing the deflection of the tip, assumed to move according to Brownian dynamics in a quadratic potential well:

$$\Delta z_{tip} = \alpha' \sqrt{24\Delta t} \sqrt{D_{tip}} + \frac{D_{tip}}{k_B T} \Delta t [-F(r) - k_{bend} \cdot z_{tip}] \quad (8)$$

Here $D_{tip} = 10D_{obst}$ is the filament tip diffusion coefficient. The true value of D_{tip} is probably greater than this, since the part of the filament free to bend is much smaller than the obstacle sizes that we consider. However, using the actual value would render the simulations extremely demanding. For this reason we have chosen a value an order of magnitude larger than D_{obst} , so that the tip fluctuations will equilibrate on time scales much faster than that of obstacle motion. The variation of the deflection z_{tip} is limited: the tip can not bend down past its own base or up so far that its height from the base exceeds the filament length. The time step $\Delta t = 10^{-9} \text{ sec}$ is chosen so that filament tips will move much less than the subunit size in one time step. The tip bending stiffness k_{bend} is obtained [36] as $k_{bend} = 3k_B T L_p / L^3 \sin^2 \theta = 0.5 \frac{pN}{nm}$, where $L_p \simeq 20 \mu m$ [37, 38] is the persistence length and L is the filament length, which we take to have a representative lamellipodium value of 100 nm. Finally α' is a random number uniformly distributed between $-\frac{1}{2}$ and $\frac{1}{2}$, so that $\langle \alpha'^2 \rangle = \frac{1}{12}$. Consecutive time steps are uncorrelated.

Previous work has calculated the time-averaged free energy of a semiflexible filament as a function of end-to-end length [39]. The present model treats the small-filament length limit of these calculations via explicit simulations. As mentioned above, time-averaged results cannot be used in the present study because the filament-tip motion must be followed with very high temporal resolution.

3. RESULTS

3.1. Thermodynamic Relation between $k_{on}(r)$ and $k_{off}(r)$

It is physically clear that at least k_{on} must be modified as a filament tip approaches an obstacle, because there is less room available for new subunits to add. Some previous models [40–43] treated hard-wall potentials and considered \bar{k}_{on} , the addition rate averaged over a time long in comparison with the time scale of filament-tip fluctuations. They argued that when the distance between the filament tip and the closest obstacle position to the filament tip in a multi-filament simulation is less than the projected length increment per subunit $\delta_{||}$, \bar{k}_{on} is reduced by a factor of $\exp[-F(\delta_{||} - r)/k_B T]$ relative to the free-filament value, where F is the time-averaged force exerted on the filament tip, and r is the distance between filament tip and the obstacle. This relationship holds when the force required to bend a filament tip is fairly constant over the size of a subunit. Here we show that a more complex relationship holds when the filament-obstacle interaction varies strongly over distances on the order of the subunit size. In such cases, it is necessary to include obstacle motion explicitly in the calculations. Then one uses rates $k_{on}(r)$ and $k_{off}(r)$ that refer to polymerization and depolymerization events occurring at a given filament-tip and obstacle position, rather than time-averaged rates.

We consider polymerization of filaments in the absence of nonequilibrium effects such as hydrolysis of ATP to ADP in actin. In this case, the stall force must be independent of the form of the interaction potential $U(r)$ between the filament tip and the obstacle. This follows from the thermodynamic arguments of Ref. [44]: At the stall force, changes

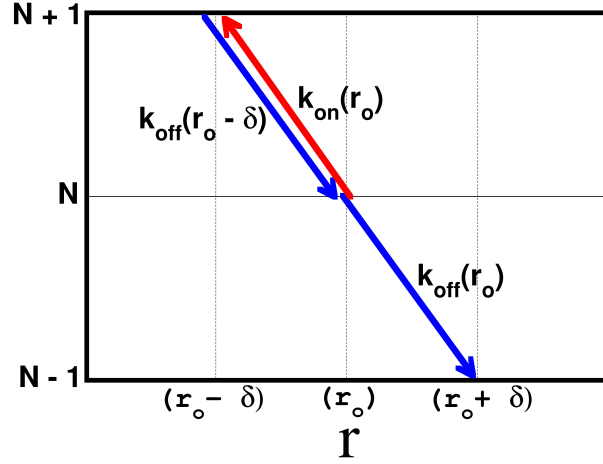


Figure 3. Dynamics of obstacle motion and actin filament polymerization and depolymerization. Red arrow corresponds to the polymerization rate at the point where the filament-obstacle gap is r_o , and the blue arrows correspond to the depolymerization rates at r_o and $r_o - \delta_{\parallel}$.

in chemical free energy resulting from polymerization precisely balance changes in mechanical energy, relating the stall force to the polymerization free energy per subunit. Then the combined system of obstacle and filament can be described by a free energy function, containing a mechanical term $F_{ext}z$ where F_{ext} is the external force (measured in the direction opposite to filament growth) acting on an obstacle with coordinate z , a chemical term $N\Delta G$ where N is the number of subunits in the filament and ΔG is the chemical free-energy increment per added subunit, and $U(r)$. Defining r to be the obstacle-tip distance, $z = r + N\delta_{\parallel}$. Then the total free energy as a function of N and r is

$$G_{tot} = N\Delta G + NF_{ext}\delta_{\parallel} + F_{ext}r + U(r). \quad (9)$$

Here the free energy is defined on a time scale shorter than the time scale of monomer addition and obstacle motion but still long enough that the free energy of an actin monomer interacting with water molecules is well defined.

The stall force F_{stall} is defined by G_{tot} being independent of N at a fixed value of r , so that

$$F_{stall} = -\Delta G/\delta_{\parallel}, \quad (10)$$

as is well known [9, 44]. This implies the well-known result [44] that

$$F_{stall} = (k_B T/\delta_{\parallel}) \ln(k_{on}^0/k_{off}^0), \quad (11)$$

where k_{on}^0 and k_{off}^0 are rates for a free filament not interacting with an obstacle. Note that k_{on}^0 is the on-rate (having units of s^{-1}), which is the product of the on-rate constant with the free-actin monomer concentration.

Now consider the dynamics of the filament-obstacle system at the stall force. The system is in equilibrium and thus obeys detailed balance (recall that ATP hydrolysis is neglected). Since the system is at the stall force, the free energy

$$G_{tot} = F_{stall} \cdot r + U(r), \quad (12)$$

is independent of N . The dynamic processes in the system are i) Brownian motion of the obstacle, ii) polymerization, and iii) depolymerization. In Figure 3, Brownian motion leads to infinitesimal steps in r , while polymerization leads to jumps $r \rightarrow r - \delta_{\parallel}$ and depolymerization leads to jumps $r \rightarrow r + \delta_{\parallel}$. Because the system is in equilibrium, the probability distribution $P(r; N)$ satisfies the Boltzmann relation. In particular, referring to Figure 3 and ignoring normalization of P ,

$$\begin{aligned} P(r; N) &= \exp\{-[U(r) + F_{stall} \cdot r]/k_B T\} \\ P(r - \delta_{\parallel}; N + 1) &= \exp\{-[U(r - \delta_{\parallel}) + F_{stall} \cdot (r - \delta_{\parallel})]/k_B T\} \end{aligned} \quad (13)$$

Detailed balance between the states $(r; N)$ and $(r - \delta_{\parallel}; N + 1)$, which holds for this equilibrium system, implies that

$$k_{on}(r)P(r; N) = k_{off}(r - \delta_{\parallel})P(r - \delta_{\parallel}; N + 1) \quad (14)$$

Therefore, using Eq. 11,

$$\begin{aligned} \frac{k_{on}(r)}{k_{off}(r - \delta_{\parallel})} &= \exp \{ -[U(r - \delta_{\parallel}) - U(r)]/k_B T \} \cdot \exp (F_{stall} \cdot \delta_{\parallel}/k_B T) \\ &= \exp \{ -[U(r - \delta_{\parallel}) - U(r)]/k_B T \} \cdot \frac{k_{on}^0}{k_{off}^0} \end{aligned} \quad (15)$$

This relationship guarantees that the correct stall force is obtained. It generalizes the well-known result [44] for the average rates that

$$\bar{k}_{on}/\bar{k}_{off} = \exp (-F\delta_{\parallel}/k_B T) k_{on}^0/k_{off}^0. \quad (16)$$

Eq. 16 follows from Eq. 15 in the limit $D_{obst} \rightarrow \infty$. To see this, note that from Eq. 15 the rate constants have the form

$$\begin{aligned} k_{off}(r) &= k_{off}^0 f(r) \\ k_{on}(r) &= k_{on}^0 \exp \{ -[U(r - \delta_{\parallel}) - U(r)]/k_B T \} f(r - \delta_{\parallel}) \end{aligned} \quad (17)$$

where f is a function of r . When $D_{obst} \rightarrow \infty$, the obstacle position distribution $P(r)$ has its thermal equilibrium form $P(r) = \exp \{ -[U(r) + Fr]/k_B T \} / Z$, where $Z = \int_{-\infty}^{\infty} \exp \{ -[U(r) + Fr]/k_B T \} dr$. Then

$$\begin{aligned} \bar{k}_{on} &= (k_{on}^0/Z) \int_{-\infty}^{\infty} \exp \{ -[U(r - \delta_{\parallel}) - U(r)]/k_B T \} \exp \{ -[U(r) + Fr]/k_B T \} f(r - \delta_{\parallel}) dr \\ &= (k_{on}^0/Z) \int_{-\infty}^{\infty} \exp \{ -[U(r - \delta_{\parallel})]/k_B T \} \exp [-Fr/k_B T] f(r - \delta_{\parallel}) dr \\ &= (k_{on}^0/Z) \exp (-F\delta_{\parallel}/k_B T) \int_{-\infty}^{\infty} \exp [-U(r)/k_B T] \exp [-Fr/k_B T] f(r) dr \end{aligned} \quad (18)$$

while

$$\bar{k}_{off} = (k_{off}^0/Z) \int_{-\infty}^{\infty} \exp [-U(r)/k_B T] \exp [-Fr/k_B T] f(r) dr \quad (19)$$

$$= \bar{k}_{on} [\exp (F\delta_{\parallel}/k_B T) k_{off}^0/k_{on}^0], \quad (20)$$

implying that Eq. 16 holds. When obstacle diffusion is not rapid, Eq. 16 will not necessarily hold because energy is dissipated by the obstacle drag, which is inversely proportional to D_{obst} according to the Einstein relation. This is not accounted for in the thermodynamic analysis above. The stall force, however, is unaffected by obstacle drag because the obstacle is stationary on average.

Eq. 15 correctly implies that the force-velocity relation is independent of the choice of zero for evaluating r . For example, adding a constant shift Δr to r would cause the typical positions sampled by the filament tip to move out a distance Δr from the obstacle, i. e. remaining close to the minimum of $U(r)$ if the potential has a deep well. Then the values sampled by the factor $\exp \{ -[U(r - \delta_{\parallel}) - U(r)]/k_B T \}$ will also remain the same, corresponding to the energy difference between a point at the minimum and one shifted in by Δr from the minimum. This also implies that for the repulsive potential, the force-velocity relation is independent of the prefactor A ; changes in the prefactor can be accounted for by changing the zero of the r -coordinate, which does not affect the force-velocity relation. This is confirmed by our numerical simulations below.

The derivation above applies to a single-stranded filament. However, Eq. 15 holds for a broader range of models. For multistranded filament growth, Figure 3 applies provided that a new subunit can add only at a unique specified site (typically next to the preceding one); then δ_{\parallel} is again the projected step size per added subunit. Thus Eq. 15 holds for both these cases. It also holds when filament bending degrees of freedom are included, and for systems of many filaments. If, for example, one describes the bending of a single filament by angle θ , then Figure 3 applies to transitions occurring at a given value of θ . Because detailed balance must hold for all transitions in a system at equilibrium, Eq. 15 will still hold. In systems of many filaments, Figure 3 would apply to a single filament, and again transitions involving just that filament must satisfy detailed balance at the stall force.

Eq. 15 does not uniquely determine $k_{on}(r)$ and $k_{off}(r)$. In our simulations, we make a minimal assumption, that

has frequently been used for the rates \bar{k}_{on} and \bar{k}_{off} , by preventing both rates from exceeding the free filament on and off rates:

$$\begin{aligned} k_{on}(r) &= k_{on}^0 \exp \{ -[U(r - \delta_{\parallel}) - U(r)]/k_B T \} & \text{if } U(r - \delta_{\parallel}) > U(r) \\ k_{on}(r) &= k_{on}^0 & \text{if } U(r - \delta_{\parallel}) < U(r) \end{aligned} \quad (21)$$

$$\begin{aligned} k_{off}(r) &= k_{off}^0 \exp \{ -[U(r + \delta_{\parallel}) - U(r)]/k_B T \} & \text{if } U(r + \delta_{\parallel}) > U(r) \\ k_{off}(r) &= k_{off}^0 & \text{if } U(r + \delta_{\parallel}) < U(r) \end{aligned} \quad (22)$$

Thus if $U(r)$ is monotonically repulsive, there is no correction to k_{off} in Eq. 22. This assumption has been made in most previous calculations in the literature. We do not have strong arguments justifying the assumption, but have decided to make it here in order to avoid investigating an unwieldy set of possibilities.

In the limit of a hard obstacle, where $U(r - \delta_{\parallel})$ jumps suddenly from 0 to ∞ when r becomes less than δ_{\parallel} , $k_{on}(r)$ in Eq. 21 will vanish when $r < \delta_{\parallel}$ and equal k_{on}^0 otherwise, as in the BR analysis. For a slowly varying repulsive $U(r)$, force balance on the obstacle implies that typical values of r will satisfy $dU/dr \simeq -F_{ext}$. Then $U(r - \delta_{\parallel}) - U(r) \simeq F_{ext}\delta_{\parallel}$, so that k_{on} is reduced by the familiar $\exp(-F_{ext}\delta_{\parallel}/k_B T)$ factor. However, in the case of an interaction potential with a deep narrow well, the results can be quite different. For vanishing external force on the obstacle, basing the slowing on the average obstacle force will give no correction. However, Eqs. 21 and 22 will give corrections to both k_{on} and k_{off} . The filament tip will generally be near the bottom of the well. Therefore both $U(r - \delta_{\parallel}) - U(r)$ and $U(r + \delta_{\parallel}) - U(r)$ are positive, so that k_{on} and k_{off} are reduced.

3.2. Numerical results for different potentials and finite D_{obst}

The instantaneous rates given in Eqs. 21 and 22 do not give the time-averaged force-velocity relation directly because the distribution of r is not known *a priori*. It is rather determined in complex ways by the combination of obstacle motion, filament-tip motion, and polymerization. Here we use simulation and analytic theory to calculate the force-velocity relations for a range of filament-obstacle interaction potentials described above. We treat “pusher” and “single-well” potentials (Figure 2a), and potentials having positive or negative Gaussian spikes as well as a double-well potential (Figure 2b). The key parameter values are given in Table I. The free-filament polymerization rate k_{on}^0 is taken for a concentration of $1\mu M$ actin with an on-rate constant of $11.6\mu M^{-1}s^{-1}$ [45]. Because the fractional error of the off-rate k_{off}^0 measured in Ref. [45] is much larger than that of the on-rate constant, we have assigned it a rough estimate of $1s^{-1}$ corresponding to the general range of values in the literature. The obstacle diffusion coefficient D_{obst} is taken as that of a sphere of radius $R = 5\mu m$, using the Stokes relation $D_{obst} = k_B T / 6\pi\mu R$, where the viscosity μ is taken as that of cytoplasm, assumed to have a value $8.9 \times 10^{-3} Pa \cdot s$ ten times larger than that of water. This corresponds to relatively rapid diffusion according to the measure $D_{obst}/k_{on}^0 \delta^2 \simeq 50$ [9]. Because this ratio is the key

TABLE I: Symbol definitions and parameter values.

Symbol	Definition	Value
δ	Actin step size	2.7 nm
D_{obst}	Obstacle diffusion coefficient	5000 nm ² /s
D_{tip}	Filament tip diffusion coefficient	5×10^4 nm ² /s
Δt	Simulation timestep	10^{-8} s
k_{on}^0	Free filament polymerization rate	$11.6 s^{-1}$
k_{off}^0	Free filament depolymerization rate	$1 s^{-1}$
F_{stall}	Filament stall force	3.74 pN
k_{bend}	Filament tip bending stiffness	0.5 pN/nm
$U(r)$	Potential of interaction between filament tip and obstacle	varies
$F(r)$	Force exerted on obstacle by filament	varies
F_{ext}	External force on obstacle	varies
v_{growth}	Filament growth velocity	varies
r	Gap between filament tip and obstacle	varies
z_{obst}	Obstacle z coordinate (height)	varies
Δz_{tip}	Filament tip fluctuation in a timestep	varies

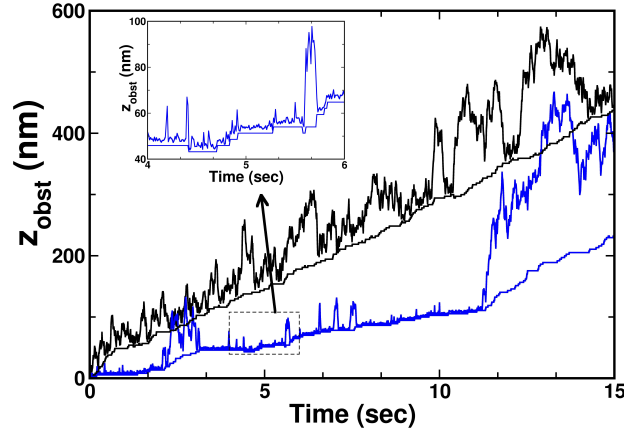


Figure 4. Time course of filament length and obstacle motion for zero external force. Upper black curve is obstacle motion time course for a repulsive filament-obstacle interaction potential (corresponding parameter values from Eq. 3 are: $A=54.7 \text{ pN}\cdot\text{nm}$, $B=0$, and $\kappa_1=0.9 \text{ nm}^{-1}$) and the lower black curve is the filament height vs. time. Upper blue curve is the time course of the obstacle motion for a potential with an attractive well of depth $5 k_B T$ (parameter values from Eq. 3 are: $A=71.6 \text{ pN}\cdot\text{nm}$, $B=35.8 \text{ pN}\cdot\text{nm}$, $\kappa_1=0.9 \text{ nm}^{-1}$, and $\kappa_2=0.3 \text{ nm}^{-1}$) with lower blue curve showing the filament height. Inset shows discrete polymerization and depolymerization steps, as well as the obstacle fluctuations against the filament tip.

factor controlling the polymerization behavior, our results could also be taken to describe, for example, a system with faster polymerization and faster diffusion. The calculated force-velocity relations are not sensitive to D_{obst} provided diffusion is much more rapid than polymerization, so they apply to a variety of obstacle sizes. The effects of using lower values of D_{obst} are described in Appendix B.

Figure 4 shows sample time courses of filament length and obstacle position at zero load for two different force fields. The black curves correspond to a “soft” repulsive potential (see Figure 2a). The filament grows at roughly the free-filament rate and the obstacle has excursions of 100 nm or more away from the filament tip. The blue curves correspond to a force field with an attractive well (Figure 2a) of depth $5 k_B T$. Here the growth is slower by about 50%. The obstacle excursions are smaller on average. Although there are several “mini-excursions” of tens of nm, the obstacle returns to the filament. We consider this case to correspond to transient attachment of the filament to the obstacle. However, later in the time course (at around 11 sec), the obstacle has an excursion similar to that seen for repulsive potential. Eventually, the filament will catch up to the obstacle and the excursions will diminish.

Figure 5 shows the calculated force-velocity relations for the range of potentials considered. For all the potentials, the growth velocity lies at or below the BR prediction. This is expected from the assumption (Eqs. 21 and 22) that $k_{off} \leq k_{off}^0$: Eq. 16 implies that the time-averaged rates $\bar{k}_{on} - \bar{k}_{off} = k_{on}^0 \exp(-F\delta/k_B T) f(F) - k_{off}^0 f(F)$, where $f(F) \leq 1$. Then $\bar{k}_{on} - \bar{k}_{off} = f(F)[k_{on}^0 \exp(-F\delta/k_B T) - k_{off}^0]$, below the BR prediction. For all purely repulsive potentials, the force-velocity relation is essentially indistinguishable from the BR prediction. This again follows from Eq. 16, 21, and 22. For repulsive potentials Eqs. 21 and 22 imply that $k_{off}(r) = \bar{k}_{off} = k_{off}(0)$, so that $\bar{k}_{on} = k_{on}^0 \exp(-F\delta/k_B T)$ and the BR relation holds.

Simple-well potentials, as well as potentials having negative spikes and double wells (Figure 2b), lead to slowing of growth relative to the BR model in the positive (pushing) force regime. For a well depth of $5 k_B T$, the zero-force velocity is about half of the free-filament velocity, larger than might have been expected from the $\exp\{-[U(r-\delta) - U(r)]/k_B T\}$ factor in Eq. 21, which is approximately $\exp(-5)$ in the case of the Gaussian spike when the obstacle is near the bottom of the potential well. The reason for the faster growth is that in the absence of external force, the obstacle is outside the well a substantial fraction of the time.

To see this, we assume fast obstacle diffusion and exploit the fact that the growth of the filament tip at velocity v toward the obstacle has the same effect on the distance distribution $P(r)$ as a weak external effective force $F_{eff} = k_B T v_{growth}/D_{obst}$ [9]. This holds because the equation of motion for $P(r)$ in the presence of filament growth (taken to have a constant velocity, and using the Einstein relation) is $\partial P/\partial t = D_{obst} \partial^2 P/\partial r^2 + [F(r)D_{obst}/k_B T + v_{growth}] \partial P/\partial r$, while the equation of motion in the presence of a constant force F_{eff} is $\partial P/\partial t = D_{obst} \partial^2 P/\partial r^2 + \{[F(r) + F_{eff}]D_{obst}/k_B T\} \partial P/\partial r$.

We assume the obstacle to be in either the region of the potential well, where we ignore polymerization, or in

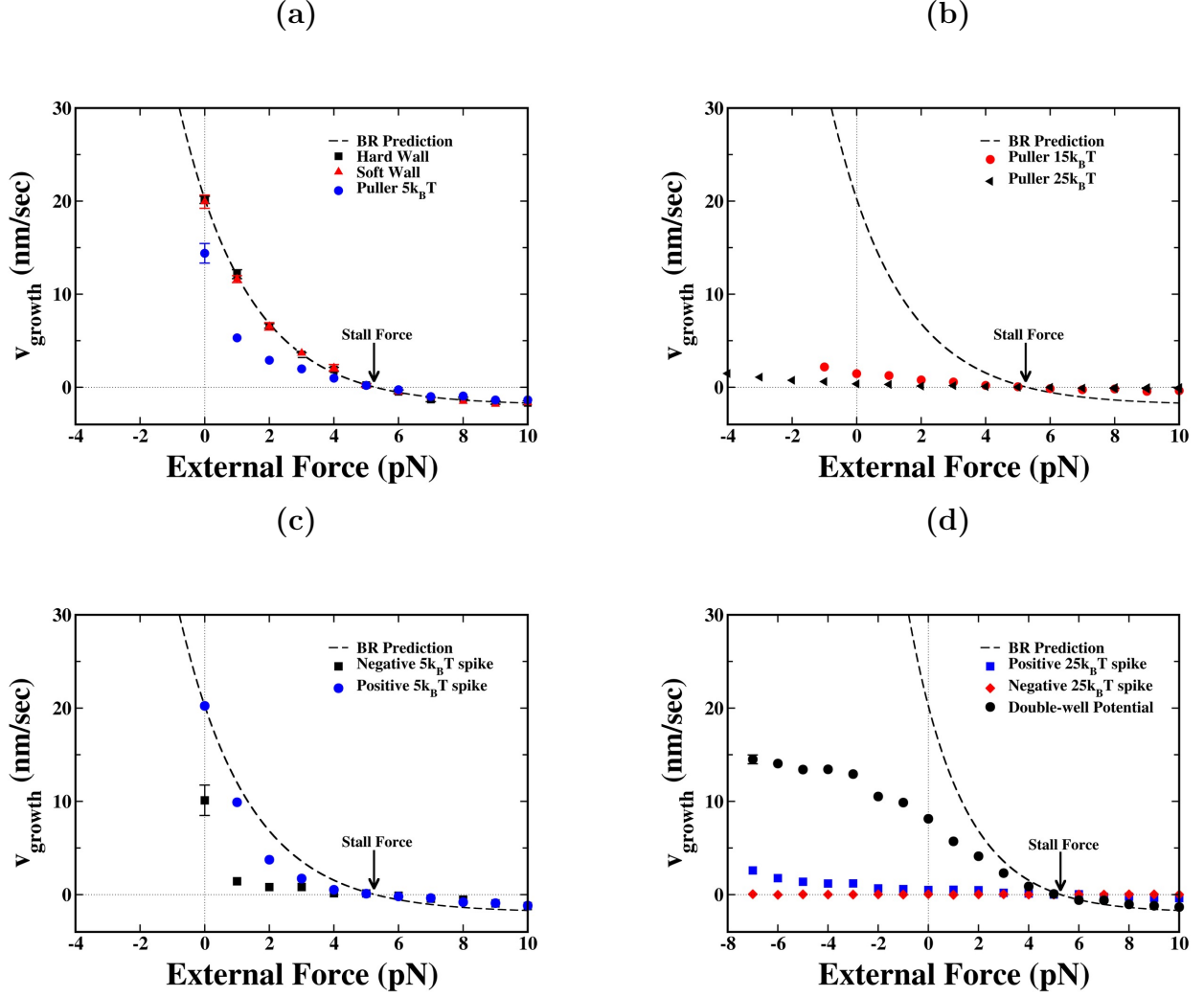


Figure 5. Simulated force-velocity relation for several different forms of filament-obstacle interaction potential. Error bars are smaller than the symbols, except indicated; and they are calculated by finding the standard deviation of the mean for growth velocity, from 20 different simulations of 20 seconds. The dashed line is the prediction of the BR model at large diffusion constant values for obstacle. Data points for pullers are shown out to forces where they detach from the obstacle. Frames (a) and (b) are for potentials in Figure 2a, while (c) and (d) are for potentials in Figure 2b.

the region outside the well, where it polymerizes at the free-filament growth velocity v_0 . This picture is most applicable to narrow wells, such as the “Negative Spike” treated in Figure 5c. The polymerization rate is then $v_{growth}/v_0 = Z_{free}/(Z_{well} + Z_{free})$, where Z_{well} is the contribution to the obstacle’s partition function from the well region and Z_{free} is the contribution from outside the well. We take

$$Z_{free} = \int_0^\infty \exp(-F_{eff} \cdot r/k_B T) dr = \frac{k_B T}{F_{eff}} = \frac{D_{obst}}{v_{growth}} \quad (23)$$

where we have taken the integral to extend from 0 to ∞ for mathematical simplicity, which is valid as long as F_{eff} is weak. This gives

$$\frac{v_{growth}}{v_0} = \frac{1}{1 + Z_{well}/Z_{free}} = \frac{1}{1 + Z_{well}v_{growth}/D_{obst}} \quad (24)$$

The solution to this equation is

$$\frac{v_{growth}}{v_0} = \frac{2}{1 + \sqrt{1 + 4\eta}}, \quad (25)$$

where $\eta = v_0 Z_{well}/D_{obst}$. Treating the “Negative $5k_B T$ Spike” as a square well of depth $5k_B T$ and width 2 nm, we obtain $Z_{well} = 2 \text{ nm} \cdot e^5$ and $v_{growth}/v_0 = 0.52$, roughly consistent with the numerical results in Figure 5c. The growth velocity given by Eq. 25 is appreciable only if $\eta \lesssim 1$, so that $D_{obst} \gtrsim v_0 Z_{well}$. Therefore, even though $D_{obst}/k_{on}^0 \delta^2 \gg 1$, the growth velocity depends strongly on the diffusion coefficient.

For the potentials with $5k_B T$ wells, the velocity also decays more rapidly with opposing force than the BR relation predicts. For example, the “ $5 k_B T$ ” curve in Figure 5a drops by nearly 75% already at 1 pN force, while the BR model and repulsive force fields drop by only about 50%. For the “Negative $5 k_B T$ ” spike in Figure 5c, the drop is practically down to zero. This rapid drop occurs because the external force reduces the statistical weight of the free region, which becomes $Z_{free} = k_B T/(F_{eff} + F_{ext})$. Substituting this into Eq. 24 gives

$$\frac{v_{growth}}{v_0} = \frac{2}{1 + \tilde{f} + \sqrt{(1 + \tilde{f})^2 + 4\eta}}, \quad (26)$$

where $\tilde{f} = Z_{well} F_{ext}/k_B T$. Even small values of F_{ext} can affect the velocity strongly, because of the Z_{well} factor in \tilde{f} . Solving Eq. 26, again using a square well of depth $5k_B T$ and width 2 nm, we find $v_{growth}/v_0 = 0.014$ at $F_{ext} = 1$ pN, consistent with the very rapid drop seen in Figure 5c. When diffusion is very rapid, $\eta \rightarrow 0$, and

$$v_{growth}/v_0 \simeq 1/(1 + \tilde{f}). \quad (27)$$

Taking $(1/v_{growth})(-dv_{growth}/dF_{ext}) \geq 2\delta/k_B T$ at zero force (twice the BR value) as a definition of rapid decay with force, we find that rapid decay will occur when $Z_{well} \gtrsim 6 \text{ nm}$.

For the double-well potential (Figure 5d), the velocity decays less rapidly relative to its $F = 0$ value than the BR prediction (although the magnitude of v_{growth} is always smaller than the BR result); in fact the drop is almost linear between 0 and 4 pN.

Another feature of the systems with potential wells is that they can polymerize processively under pulling (negative) force, over a limited time [27]. As expected physically, there is a tradeoff between maximum sustainable force and polymerization rate, illustrated in Fig. 6. High pulling force enhances polymerization, but at the same time accelerates detachment of the obstacle from the filament. We consider 20 sec as a typical time frame for a cytoskeletal process such as generation of an endocytic invagination. The results in Fig. 6 will change if different time scales are considered. However, because the detachment rate depends exponentially on the pulling force, the pulling force will conversely have a slow, logarithmic dependence on the time scale. For a $5k_B T$ potential depth, almost no pulling force can be sustained over 20 seconds (Figure 5a). But for a $15k_B T$ depth (Figure 5b), a force of about 1 pN can be sustained. The growth velocity in this case is about 80% greater than the zero-force value, but much lower than for the $5 k_B T$ case. For the $25k_B T$ potential still larger pulling forces can be sustained, at an even smaller growth rate. Figs. 5(c) and Fig. 5d show that the same trade-off occurs for potentials with spikes. In Figure 6, frame a) shows how the maximum pulling force that allows a 20-second attachment period, depends on the depth of the potential well. Frame b) shows how the growth velocity depends on the well depth. These two plots show that large sustainable pulling forces on the obstacle (greater than about 1 pN) come at the expense of greatly reduced growth velocity. Pullers with a well depth of about $5k_B T$ grows only half as fast as a free filament, but can sustain essentially no pulling force. If the depth is larger than about $15k_B T$, which is needed to sustain pN forces, growth almost completely stops for the potential widths used here. For broader potential wells, the tradeoff between velocity and maximum pulling force will be less stringent, because the exponential reduction of k_{on} in Eq. 17 will be less pronounced, allowing more rapid growth at a given potential depth. The double-well potential (Figure 2b) gives an example of this. As shown in Figure 5d, this potential eliminates the trade-off between sustainable force and polymerization rate that is seen with other potentials. This is because the well is wide enough that polymerization can occur inside even a very deep well. If the obstacle is pulled toward the large- r end of the well, $U(r - \delta)$ and $U(r)$ do not differ greatly, so there is no significant slowing in Eq. 21.

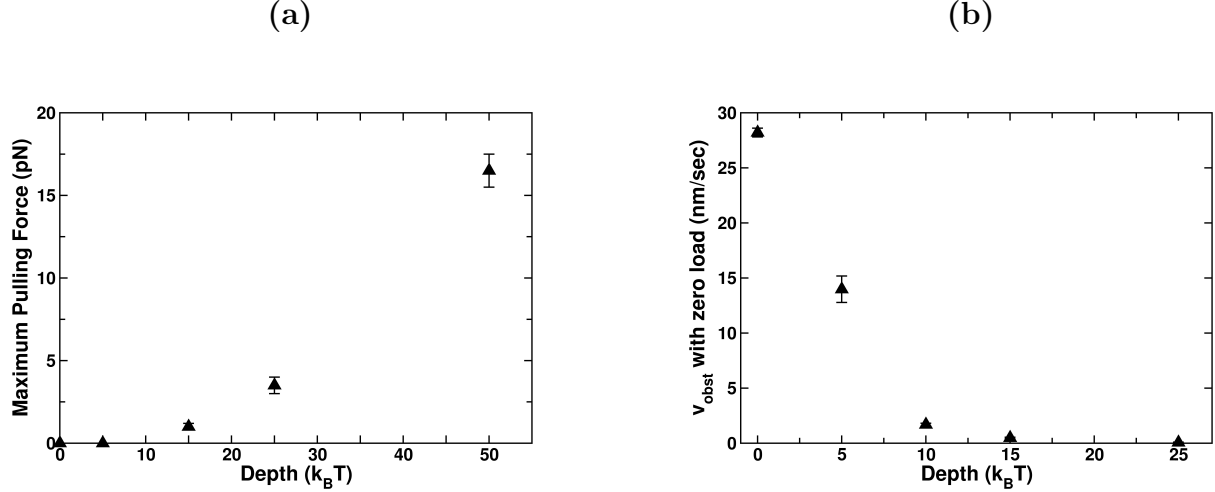


Figure 6. Effect of binding strength of single-well potential on a) the maximum pulling force allowing attachment up to 20 seconds, and b) the growth velocity at zero external load.

4. DISCUSSION

Our main findings are the following:

4.1. Thermodynamically consistent on- and off-rates

The ratio of the on- and off-rates depends on the details of the interaction potential with the obstacle on a nanometer scale (see Eq. 15). This implies that even at zero force, a deep well in the interaction potential can dramatically slow both polymerization and depolymerization. Polymerization is slowed because of the high-energy configuration assumed by the filament tip after addition of a new subunit, while depolymerization is slowed because removal of an existing subunit also leaves the filament in a high-energy configuration. Eq. 15 is automatically satisfied by calculations implementing the BR approximation that $k_{on}(r) = 0$ when the filament tip is within a distance δ of a hard obstacle with no attractive well, but $k_{on}(r) = k_{on}^0$ beyond that distance.

Not all single-filament calculations in the literature treating obstacle motion explicitly have satisfied the constraint of Eq. 15. We note that Eq. 15 does not apply to calculations such as those of Refs. [20], which ignored depolymerization. Ref. [22] assumed a variation of the on-rate resulting from the interaction potential with the obstacle, but it did not appear to satisfy Eq. 15. On the other hand, Ref. [27] used a relationship very similar to Eq. 15, in which the exponential factor corresponded to the energy difference between binding in two different positions in a double-well potential.

Some works treating force generation by actin networks stochastically have also treated smoothly varying tip-obstacle interactions [4, 46–51]. In all of these works, actin polymerization was slowed by the obstacle. But none of the treatments satisfies Eq. 15 exactly. Ref. [46] used an approximate version in which the $U(r)$ term in the exponential of Eq. 15 is ignored. In Refs. [47] and [48], the mechanical energy, including the membrane deformation/position and states of actin filament bending, was minimized after each polymerization event. To evaluate the on-rate, the energy was reminimized with a hypothetical next subunit in place. The resulting energy difference ΔU was used in a Boltzmann factor slowing the polymerization. This approach is similar in spirit to Eq. 15, but differs in that the energy is minimized in calculating ΔU . In the context of the systems treated here, this would correspond to placing the obstacle, before and after the addition of the new subunit, at the minimum of the potential $U(r) + F \cdot r$. Then $\Delta U = F\delta_{\parallel}$, giving a slowing of the growth velocity of a single filament by a factor of $\exp(-F\delta_{\parallel}/k_B T)$. As found above, this is correct provided there is no attractive well in the potential.

Refs. [4, 49–51] used criteria based on force rather than energy difference. In such approaches, even plausible assumptions regarding the slowing of actin polymerization can lead to substantial errors in the stall force. For example, one can assume [4] that a filament tip experiences a linear force $F(r) = -k_c r \theta(-r)$ when in contact with

the obstacle, and that this force slows polymerization according to

$$k_{on}(r) = k_{on}^0 \exp[-F(r)\delta_{\parallel}/k_B T], \quad (28)$$

while $k_{off}(r) = k_{off}^0$ is not affected. This corresponds to applying the thermodynamic result Eq. 16, for the averaged rate \bar{k}_{on} , to the instantaneous rate $k_{on}(r)$. The force-velocity relation can be obtained exactly for this model in the limit $D_{obst} \rightarrow \infty$. Using the same analysis as used to derive Eq. 18, but incorporating Eq. 28, we obtain

$$\begin{aligned} \bar{k}_{on} &= (k_{on}^0/Z) \int_{-\infty}^{\infty} \exp[-F(r)\delta_{\parallel}/k_B T] \exp\{-[U(r) + F_{ext} \cdot r]/k_B T\} dr \\ &= (k_{on}^0/Z) \left[\sqrt{\frac{\pi k_B T}{2k_c}} \exp\{[F_{ext} - k_c \delta_{\parallel}]^2/2k_c k_B T\} \{1 + \text{erf}[(F_{ext} - k_c \delta_{\parallel})/\sqrt{(2k_c k_B T)}]\} + k_B T/F_{ext} \right] \end{aligned} \quad (29)$$

where $U(r) = k_c \theta(-r)r^2/2$ and

$$Z = \sqrt{\frac{\pi k_B T}{2k_c}} \exp(F_{ext}^2/2k_c k_B T) \{1 + \text{erf}(F_{ext}/\sqrt{(2k_c k_B T)})\} + k_B T/F_{ext}. \quad (30)$$

The stall force for this model can be obtained by numerically locating the zero of $\bar{k}_{on} - \bar{k}_{off}$.

In the limit of large stall force ($F_{ext} - k_c \delta_{\parallel} \gg \sqrt{2k_c k_B T}$), Eq. 29 simplifies. The error functions in Eqs. 29 and 30 approach unity, and the $1/F_{ext}$ terms can be ignored. Thus $\bar{k}_{on} \simeq k_{on}^0 \exp(-F_{ext}\delta_{\parallel}/k_B T) \exp(k_c \delta_{\parallel}^2/2k_B T)$. Solving for $\bar{k}_{on} = \bar{k}_{off}$ we obtain the incorrect result

$$F_{stall} = (k_B T/\delta_{\parallel}) \ln(k_{on}^0/k_{off}^0) + k_c \delta_{\parallel}/2, \quad (31)$$

which exceeds the thermodynamically derived stall force by the amount $k_c \delta_{\parallel}/2$. Numerically, both from Eq. 29 and from our stochastic simulations with a large but finite D_{obst} , we find comparable overestimates of the stall force in other parameter ranges. The overestimates are equally large in parameter ranges where filament-tip fluctuations dominate. For the spring constant assumed in Ref. [4] we obtain a stall force of over 50 pN for a filament at 45° incidence, several times too high given the assumed actin concentration. We emphasize that these results hold only when thermal motion of the obstacle or filament tip is explicitly treated rather than being averaged out.

Why does Eq. 29, which appears to correctly implement the force dependence of actin polymerization by equating the instantaneous rates to the known results (Eq. 16) for the average rates, fail to obtain the correct stall force? The reason is that although the force $F(t)$ acting on the tip of a filament polymerizing under an average force F_{ext} satisfies $\bar{F} = F_{ext}$, the fluctuating motion of the obstacle or the filament tip causes the tip to experience a range of forces described by a distribution $p(F)$, so that

$$\bar{k}_{on} = k_{on}^0 \int_{-\infty}^{\infty} p(F) \exp(-F\delta_{\parallel}/k_B T) dF \quad (32)$$

Since the exponential is convex upwards (the second derivative of $\exp(-F\delta_{\parallel}/k_B T)$ is positive), \bar{k}_{on} will exceed $k_{on}(\bar{F})$, leading to an overestimate of the stall force. If one assumes that $p(F) \propto \exp[-(F - \bar{F})^2/2\Delta F^2]$ where ΔF^2 is the variance of F , Eq. 32 shows that $\bar{k}_{on} = k_{on}^0 \exp[-\bar{F}\delta_{\parallel}/k_B T] \exp[\delta_{\parallel}^2 \Delta F^2/2(k_B T)^2]$. Setting this equal to k_{off}^0 , one finds the following overestimate of the stall force:

$$F_{stall} = (k_B T/\delta_{\parallel}) \ln(k_{on}^0/k_{off}^0) + \Delta F^2 \delta_{\parallel}/2k_B T. \quad (33)$$

For force fluctuations induced by thermal motion in a harmonic potential, $\Delta F^2 = \Delta r^2 k_c^2 = k_c k_B T$, and Eq. 33 is identical to Eq. 31. On the other hand, if thermal motions of the filament tip or obstacle come from non-thermal sources, as in Refs. [49–51], force fluctuations could instead result from the geometrical constraints imposed on the individual filaments by the network structure. Then one might expect the bending-induced fluctuation of a given filament tip position to be $\lesssim \delta_{\parallel}$; larger deformations would likely be evened out by the differences in growth velocity. As described in Appendix A, one can estimate the spring constant of a filament in the direction of motion as $k_{bend} \simeq 0.5 \frac{pN}{nm}$. Then at 45° incidence $\Delta F \sim k_{bend} \delta_{\parallel} \lesssim 0.92$ pN, and the overestimate in Eq. 33 is $\lesssim 0.3$ pN, much smaller than for thermal force fluctuations.

4.2. Very rapid decay of force-velocity relation for some potentials

For relatively shallow attractive potentials the velocity can be a substantial fraction of the free-filament velocity at zero force, but decay rapidly with opposing force. This finding may help explain the results of experiments [52] studying small number of actin filaments growing against a hard wall, if the filament tips are weakly bound to the wall. The filaments propelled acrosomes attached to beads held in an optical trap (backwards). The filaments/acrosome/bead are moving rather than the obstacle, so as discussed above it is their motion that is considered. The growth velocity was found to drop off much more rapidly than expected from the BR model at forces of a few tenths of a pN per filament, especially at a $2\mu M$ actin concentration. We cannot treat their many-filament system within our model, so we consider a single filament growing against an obstacle with a force of a few tenths of a pN . Because the bead is trapped in potential well of spring constant $k_c \simeq 0.008 pN/nm$, the “free” contribution Z_{free} is different from that calculated in Sec. 3.2. The energy of the bead in the potential well, displaced a distance r from the minimum, is $k_c r^2/2$. Thus $Z_{free} = \int_0^\infty \exp(-F_{ext} \cdot r) \exp(-k_c r^2/2) dr$. Provided that $F_{ext} \geq \sqrt{k_c k_B T} = 0.2 pN$, the reduction in Z_{free} from F_{ext} will exceed that from k_c , and it is reasonable to take $Z_{free} = k_B T / F_{ext}$ as in Sec. 3.2. Then Eq. 27 applies, and a force of few tenths of a pN per filament could reduce the velocity by a large factor if $Z_{well} \geq 15 nm$. The validity of Eq. 27 requires that $\eta = v_0 Z_{well} / D_{obst} < 1$. In Ref. [52], v_0 was less than $20 nm/s$, so a diffusion coefficient $\geq 600 nm^2/s$ would be adequate.

Experiments on whole cells [19, 53] have shown that the growth velocity of lamellipodia drops very rapidly with opposing force. The velocity decay found here may contribute to this effect, but only if the filaments are sufficiently long to allow thermal fluctuations greater than $Z_{free} = k_B T / F_{ext}$ away from the obstacle. In this system, mechanical factors may be the dominant effect [19].

4.3. Upper bound on growth rate

Provided that $k_{off}(r) \leq k_{off}^0$ and D_{obst} is large, no type of filament-obstacle interaction potential leads to polymerization faster than a hard wall; for monotonically decaying force fields the force-velocity relation is very near that for a hard wall, which is described well by the BR prediction. These results follow from Eq. 16 and are confirmed by the simulations. For intermediate values of $D_{obst} \sim k_{on}^0 \delta_{||}^2$, we find that softer potentials accelerate growth slightly (see Figure B.1).

But when D_{obst} is very small, a soft potential can accelerate polymerization substantially. This was found in the simulations of [22], where a soft obstacle accelerated polymerization by about 100% using $D_{obst} = 0.0016 k_{on}^0 \delta^2$ for a filament growing at perpendicular incidence so that $\delta_{||} = \delta$. To understand this physically, consider the case of zero external force. In the limit of a slowly varying potential, it is legitimate to ignore the randomness in polymerization and treat filament growth as occurring at a constant velocity. As described in Section 3.2, obstacle drag can then be included via an effective force $F_{eff} = k_B T v_{obst} / D_{obst}$ [9]. After a sufficiently long time, the probability distribution $P(r)$ will settle into a steady-state form $P(r) = \exp\{-[U(r) + F_{eff} \cdot r] / k_B T\} / Z$, where Z is the corresponding partition function. Then following the derivation of Eq. 18, and taking $f(r) = 1$ in Eq. 17 so that $\bar{k}_{off} = k_{off}^0$,

$$\begin{aligned} \bar{k}_{on} &= (k_{on}^0 / Z) \exp(-F_{eff} \delta / k_B T) \int_{-\infty}^{\infty} \exp[-U(r) / k_B T] \exp[-F_{eff} \cdot r / k_B T] dr \\ &= \exp(-F_{eff} \delta / k_B T) k_{on}^0 \end{aligned} \quad (34)$$

Since $v_{obst} = (\bar{k}_{on} - k_{off}^0) \delta$, it follows that $v_{obst} = (D_{obst} x / \delta - k_{off}^0 \delta)$, where x satisfies the transcendental equation $x / \hat{k}_{on} = \exp(-x) \exp(\hat{k}_{off})$ and we define $\hat{k}_{on, off} = (k_{on, off}^0 \delta^2 / D_{obst})$. Therefore

$$v_{obst} = (D_{obst} / \delta) \{W[\hat{k}_{on} \exp(\hat{k}_{off})] - \hat{k}_{off}\} \quad (35)$$

$$\simeq (D_{obst} / \delta) \ln(\hat{k}_{on} / \hat{k}_{off}) \quad (\hat{k}_{on} \gg 1, \hat{k}_{off} \gg \ln(\hat{k}_{on})) \quad (36)$$

$$\simeq (D_{obst} / \delta) [\ln(\hat{k}_{on}) - \ln(\ln \hat{k}_{on})] \quad (\hat{k}_{on} \gg 1, \hat{k}_{off} \rightarrow 0) \quad (37)$$

where the Lambert function W is the inverse of the function $x \exp(x)$. We have used the asymptotic expansion $W(x) \simeq \ln(x) - \ln[\ln(x)]$, valid when x is large (very slow diffusion).

In cases of very slow obstacle diffusion, the simplified form Eq. 36 will hold if the ratio between the on- and off-rates is moderate. The physical content of this result is clarified by considering the work done in moving the obstacle. The work done per added subunit is $(v_{obst} k_B T / D_{obst}) \delta$ where the first term is the force according to the Einstein relation. From Eq. 36 this equals $k_B T \ln(k_{on}^0 / k_{off}^0)$, which is the free energy released per added subunit [44]. Thus the process

is 100% efficient in that all of the free energy of polymerization is used to push the obstacle against the drag force. If (k_{on}^0/k_{off}^0) is large, the velocity in Eq. 36 could exceed the hard-wall limit $2D_{obst}/\delta$ [9] substantially. For example, for a free-actin concentration of $10\mu M$, our parameters would predict that $(k_{on}^0/k_{off}^0) \simeq 100$, and the logarithm is greater than 4. Then the hard-wall limit would be exceeded by more than a factor of 2.

The simulations of Ref. [22] treated the case $\hat{k}_{on} = 6.67 \times 10^2$ and $\hat{k}_{off} = 0$. In this case, Eq. 35 gives $v_{obst} = 4.9D_{obst}/\delta$, while Eq. 37 gives $v_{obst} = 4.6D_{obst}/\delta$, both about a factor of two above the hard-wall limit and comparable with the value of $4.4D_{obst}/\delta$ found in Ref. [22]. This suggests that the analytic theory captures the key effects in these simulations.

4.4. Requirements for coexistence of strong pulling forces and polymerization

Sustaining strong pulling forces at a significant rate of polymerization requires a deep, broad well in the filament-obstacle interaction. Actin filaments polymerizing under pulling force may have several functions. They could act as parts of force sensors, or as mechanical absorbers for rapidly generated forces from myosin motors [2]. It is also believed that actin filaments in the central region of endocytic sites in budding yeast exert pulling forces on the membrane [4, 6], and it is important to know if filaments can polymerize sufficiently quickly to generate a gel in the pulling region that can sustain the large stresses generated by the process. The behavior of actin filaments under pulling forces has been addressed by thermodynamic arguments [54] as well as kinetic models [2, 55] and simulations [27, 56]. Most of the calculations have predicted acceleration of polymerization by pulling force, but Ref. [56] found a competition between different conformational factors that could either slow or speed polymerization. Experimental studies [2, 55, 57, 58] have shown that polymerization under pulling forces of several pN is possible if actin filaments are linked to the obstacle via formins. They suggest, on the whole, that pulling force accelerates polymerization if rotational constraints are absent. These models have included effects not explicitly included here, such as conformational changes of formins at the actin filament tip. Our finding that for narrow wells polymerization is incompatible with the ability to sustain large forces implies that in the systems where this phenomenon occurs, the interaction between the actin filament and the obstacle must have a broad minimum. Our models are too simple to quantitatively describe the three-dimensional geometry of a formin-tipped actin filament, but the broad minimum in our “double-well” potential may approximately mimic the conformational flexibility that appears to be at the heart of the phenomenon.

4.5. Impact of key approximations

The calculations described here make several major approximations, including the treatment of just a single filament, the modeling of the filament-obstacle interaction via simple potential energy functions, the neglect of hydrolysis effects, and the use of single filament length to calculate fluctuations. Hydrolysis of actin filaments is a two-step process involving cleavage of the phosphate bond and subsequent release of the phosphate group. Most of the effects of hydrolysis on polymerization/depolymerization dynamics occur only after the phosphate-release step. In *in vitro* experiments, phosphate release is slow, generally requiring several minutes or more to occur. Therefore neglect of hydrolysis effects may be valid for a broad range of *in vitro* experiments. Phosphate release *in vivo* is faster, generally believed to occur over a time scale of tens of seconds. Application of our results to *in vivo* studies will be limited to phenomena where the time for a new subunit to add is substantially less than the time scale of phosphate release. We expect the use of a single filament length to be valid provided the filaments are long enough that the filament-tip fluctuations exceed the subunit size, and short enough that the filament do not buckle. In this regime the filament-tip fluctuations are rapid enough to allow monomer intercalation even if obstacle diffusion is slow. This gives a large window of filament lengths between roughly 40 nm (derived from k_{bend} in Sec. 2.3) and the buckling length, which for a force of 1 pN is about 500 nm. In practice we find little difference between our simulation results with and without tip fluctuations, again suggesting that the precise value of the filament length is not important.

The treatment of the filament bending energy via Eq. 8, and the corresponding filament-obstacle forces, are also approximate. They have been treated more completely in Ref. [34]. To explore the validity of our approximation, we analyze the average force on the obstacle obtained by the model for a single filament in the limit of a fixed obstacle and a hard-wall filament-obstacle interaction, in the absence of polymerization. As in Ref. [34], we define ζ as the distance from the base of the filament to the obstacle. The energy function that gives the bending force in Eq. 8 is $E_{bend} = k_{bend} \cdot z_{tip}^2/2$. The possible values of z_{tip} are constrained by i) the hard obstacle, which requires that $z_{tip} < \zeta - L \cos \theta$; ii) the fact that the filament cannot extend more than a distance L in the z -direction, which implies that $z_{tip} < L(1 - \cos \theta)$; and iii) our assumption that the filament tip cannot be below the filament base, which implies that $z_{tip} > -L \cos \theta$. We define z_{max} as the smaller of $\zeta - L \cos \theta$ and $L(1 - \cos \theta)$. In the absence of polymerization,

the system properties can be calculated using equilibrium thermodynamics. We thus obtain the force from the free energy G via the partition function:

$$\begin{aligned} Z &= \int_{-L \cos \theta}^{z_{max}} \exp(-k_{bend} z_{tip}^2 / 2k_B T) dz_{tip} \\ &= (\text{const}) \times [\text{erf}(z_{max}/\delta z_0) + \text{erf}(L \cos \theta / \delta z_0)] \end{aligned} \quad (38)$$

where erf is the error function and $\delta z_0 = \sqrt{2k_B T / k_{bend}}$. The corresponding time-averaged force exerted by the filament on the obstacle is (for $\zeta < L$)

$$\begin{aligned} f_{bend} &= -\frac{\partial G}{\partial \zeta} = k_B T \frac{\partial \ln Z}{\partial \zeta} \\ &= \frac{k_B T}{\delta z_0} \frac{\exp[-(\zeta - L \cos \theta)^2 / \delta z_0^2]}{[\text{erf}(z_{max}/\delta z_0) + \text{erf}(L \cos \theta / \delta z_0)]}, \end{aligned} \quad (39)$$

where we have used $G = -k_B T \ln Z$. This force has a form quite similar to the force plots of Ref. [34], slowly rising from 0 as ζ drops from L , then increasing fairly linearly after ζ drops below the contact point $L \cos \theta$. The approach used here is certainly not quantitatively accurate, but reproduces the key physical features of the more complete calculations.

To make direct contact with either *in vitro* or *in vivo* experiments will require more complex calculations for many-filament systems. The present results can help make progress toward this goal by informing multiscale calculations such as those of Ref. [10, 12–19, 28–32, 59, 60], which treat force generation by multifilament systems using a variety of approximations to include the single-filament force-velocity relation. The present results will provide useful guidance, especially in cases where different types of filament-obstacle interactions are present in the same system.

Acknowledgments

This work was supported by the National Institute of General Medical Sciences (<https://www.nigms.nih.gov>) under Grant R01 GM107667 and the National Science Foundation (<https://www.nsf.gov>) under Grant Agreement CMMI:15-458571.

-
1. Bieling, P., S. D. Hansen, O. Akin, C. C. Hayden, D. A. Fletcher, R. D. Mullins, et al., 2017. WH2 and proline-rich domains of WASP-family proteins collaborate to accelerate actin filament elongation. *The EMBO journal* e201797039.
 2. Yu, M., X. Yuan, C. Lu, S. Le, R. Kawamura, A. K. Efremov, Z. Zhao, M. M. Kozlov, M. Sheetz, A. Bershadsky, et al., 2017. mDia1 senses both force and torque during F-actin filament polymerization. *Nature communications* 8:1650.
 3. Kaksonen, M., Y. Sun, and D. G. Drubin, 2003. A pathway for association of receptors, adaptors, and actin during endocytic internalization. *Cell* 115:475–487.
 4. Mund, M., J. A. van der Beek, J. Deschamps, S. Dmitrieff, J. L. Monster, A. Picco, F. Nédélec, M. Kaksonen, and J. Ries, 2018. Systematic Nanoscale Analysis of Endocytosis Links Efficient Vesicle Formation to Patterned Actin Nucleation. *Cell* 174:884–896.
 5. Tweten, D., P. Bayly, and A. Carlsson, 2017. Actin growth profile in clathrin-mediated endocytosis. *Physical Review E* 95:052414.
 6. Wang, X., B. J. Galletta, J. A. Cooper, and A. E. Carlsson, 2016. Actin-regulator feedback interactions during endocytosis. *Biophysical journal* 110:1430–1443.
 7. Waterman-Storer, C. M., J. Gregory, S. F. Parsons, and E. Salmon, 1995. Membrane/microtubule tip attachment complexes (TACs) allow the assembly dynamics of plus ends to push and pull membranes into tubulovesicular networks in interphase *Xenopus* egg extracts. *The Journal of Cell Biology* 130:1161–1169.
 8. Perez, F., G. S. Diamantopoulos, R. Stalder, and T. E. Kreis, 1999. CLIP-170 highlights growing microtubule ends in vivo. *Cell* 96:517–527.
 9. Peskin, C. S., G. M. Odell, and G. F. Oster, 1993. Cellular motions and thermal fluctuations: the Brownian Ratchet. *Biophysical journal* 65:316–324.
 10. Mogilner, A., and L. Edelstein-Keshet, 2002. Regulation of actin dynamics in rapidly moving cells: a quantitative analysis. *Biophysical journal* 83:1237–1258.
 11. Atilgan, E., D. Wirtz, and S. X. Sun, 2005. Morphology of the lamellipodium and organization of actin filaments at the leading edge of crawling cells. *Biophysical journal* 89:3589–3602.
 12. Mogilner, A., and B. Rubinstein, 2005. The physics of filopodial protrusion. *Biophysical journal* 89:782–795.

13. Rubinstein, B., K. Jacobson, and A. Mogilner, 2005. Multiscale two-dimensional modeling of a motile simple-shaped cell. *Multiscale Modeling & Simulation* 3:413–439.
14. Marée, A. F., A. Jilkine, A. Dawes, V. A. Grieneisen, and L. Edelstein-Keshet, 2006. Polarization and movement of keratocytes: a multiscale modelling approach. *Bulletin of mathematical biology* 68:1169–1211.
15. Ditlev, J. A., N. M. Vacanti, I. L. Novak, and L. M. Loew, 2009. An open model of actin dendritic nucleation. *Biophysical journal* 96:3529–3542.
16. Joanny, J.-F., and J. Prost, 2009. Active gels as a description of the actin-myosin cytoskeleton. *HFSP journal* 3:94–104.
17. Khamviwath, V., J. Hu, and H. G. Othmer, 2013. A continuum model of actin waves in Dictyostelium discoideum. *PLoS one* 8:e64272.
18. Camley, B. A., Y. Zhao, B. Li, H. Levine, and W.-J. Rappel, 2017. Crawling and turning in a minimal reaction-diffusion cell motility model: coupling cell shape and biochemistry. *Physical Review E* 95:012401.
19. Zimmermann, J., C. Brunner, M. Enculescu, M. Goegler, A. Ehrlicher, J. Käs, and M. Falcke, 2012. Actin filament elasticity and retrograde flow shape the force-velocity relation of motile cells. *Biophysical journal* 102:287–295.
20. Carlsson, A., 2000. Force-velocity relation for growing biopolymers. *Physical Review E* 62:7082.
21. Burroughs, N., and D. Marenduzzo, 2005. Three-dimensional dynamic Monte Carlo simulations of elastic actin-like ratchets. *The Journal of chemical physics* 123:174908.
22. Burroughs, N. J., and D. Marenduzzo, 2006. Growth of a semi-flexible polymer close to a fluctuating obstacle: application to cytoskeletal actin fibres and testing of ratchet models. *Journal of Physics: Condensed Matter* 18:S357.
23. Mogilner, A., and G. Oster, 2003. Force generation by actin polymerization II: the elastic ratchet and tethered filaments. *Biophysical journal* 84:1591–1605.
24. Banigan, E. J., K.-C. Lee, and A. J. Liu, 2013. Control of actin-based motility through localized actin binding. *Physical biology* 10:066004.
25. Dickinson, R. B., and D. L. Purich, 2002. Clamped-filament elongation model for actin-based motors. *Biophysical Journal* 82:605–617.
26. Dickinson, R. B., L. Caro, and D. L. Purich, 2004. Force generation by cytoskeletal filament end-tracking proteins. *Biophysical journal* 87:2838–2854.
27. Zhu, J., and A. Carlsson, 2006. Growth of attached actin filaments. *The European Physical Journal E: Soft Matter and Biological Physics* 21:209–222.
28. Ryan, G. L., D. Holz, S. Yamashiro, D. Taniguchi, N. Watanabe, and D. Vavylonis, 2017. Cell protrusion and retraction driven by fluctuations in actin polymerization: A two-dimensional model. *Cytoskeleton* 74:490–503.
29. Craig, E. M., D. Van Goor, P. Forscher, and A. Mogilner, 2012. Membrane tension, myosin force, and actin turnover maintain actin treadmill in the nerve growth cone. *Biophysical journal* 102:1503–1513.
30. Adler, Y., and S. Givli, 2013. Closing the loop: Lamellipodia dynamics from the perspective of front propagation. *Physical Review E* 88:042708.
31. Carlsson, A. E., and P. V. Bayly, 2014. Force generation by endocytic actin patches in budding yeast. *Biophysical journal* 106:1596–1606.
32. Barnhart, E. L., J. Allard, S. S. Lou, J. A. Theriot, and A. Mogilner, 2017. Adhesion-dependent wave generation in crawling cells. *Current Biology* 27:27–38.
33. Bieling, P., S. D. Hansen, O. Akin, C. C. Hayden, D. A. Fletcher, R. D. Mullins, et al., 2018. WH2 and proline-rich domains of WASP-family proteins collaborate to accelerate actin filament elongation. *The EMBO journal* 37:102–121.
34. Gholami, A., J. Wilhelm, and E. Frey, 2006. Entropic forces generated by grafted semiflexible polymers. *Physical Review E* 74:041803.
35. Doi, M., and S. F. Edwards, 1988. The theory of polymer dynamics, volume 73. oxford university press.
36. Lautrup, B., 2005. Physics of continuous matter. *Exotic and Everyday Phenomena in the Macroscopic World*, IOP. We evaluated Eq. (10.12) of this reference at the end of the rod ($|z| = L$), and used the relationship $EI = k_B T L_p$, to obtain the formula for k_{bend} .
37. Gittes, F., B. Mickey, J. Nettleton, and J. Howard, 1993. Flexural rigidity of microtubules and actin filaments measured from thermal fluctuations in shape. *The Journal of cell biology* 120:923–934.
38. Isambert, H., P. Venier, A. C. Maggs, A. Fattoum, R. Kassab, D. Pantaloni, and M.-F. Carlier, 1995. Flexibility of actin filaments derived from thermal fluctuations. Effect of bound nucleotide, phalloidin, and muscle regulatory proteins. *Journal of Biological Chemistry* 270:11437–11444.
39. Kroy, K., and E. Frey, 1996. Force-extension relation and plateau modulus for wormlike chains. *Physical review letters* 77:306.
40. Mogilner, A., and G. Oster, 1999. The polymerization ratchet model explains the force-velocity relation for growing microtubules. *European Biophysics Journal* 28:235–242.
41. van Doorn, G. S., C. Tănase, B. M. Mulder, and M. Dogterom, 2000. On the stall force for growing microtubules. *European Biophysics Journal* 29:2–6.
42. Son, J., G. Orkoulas, and A. B. Kolomeisky, 2005. Monte Carlo simulations of rigid biopolymer growth processes. *The Journal of chemical physics* 123:124902.
43. Krawczyk, J., and J. Kierfeld, 2011. Stall force of polymerizing microtubules and filament bundles. *EPL (Europhysics Letters)* 93:28006.
44. Hill, T. L., 2012. Linear aggregation theory in cell biology. Springer Science & Business Media.
45. Pollard, T. D., 1986. Rate constants for the reactions of ATP-and ADP-actin with the ends of actin filaments. *The Journal of cell biology* 103:2747–2754.

46. Carlsson, A. E., 2001. Growth of branched actin networks against obstacles. *Biophysical journal* 81:1907–1923.
47. Schaus, T. E., E. W. Taylor, and G. G. Borisy, 2007. Self-organization of actin filament orientation in the dendritic-nucleation/array-treadmilling model. *Proceedings of the National Academy of Sciences* 104:7086–7091.
48. Schaus, T. E., and G. G. Borisy, 2008. Performance of a population of independent filaments in lamellipodial protrusion. *Biophysical journal* 95:1393–1411.
49. Hu, L., and G. A. Papoian, 2010. Mechano-chemical feedbacks regulate actin mesh growth in lamellipodial protrusions. *Biophysical journal* 98:1375–1384.
50. Zhu, J., and A. Mogilner, 2012. Mesoscopic model of actin-based propulsion. *PLoS computational biology* 8:e1002764.
51. Hu, L., and G. A. Papoian, 2013. Molecular transport modulates the adaptive response of branched actin networks to an external force. *The Journal of Physical Chemistry B* 117:13388–13396.
52. Footer, M. J., J. W. Kerssemakers, J. A. Theriot, and M. Dogterom, 2007. Direct measurement of force generation by actin filament polymerization using an optical trap. *Proceedings of the National Academy of Sciences* 104:2181–2186.
53. Prass, M., K. Jacobson, A. Mogilner, and M. Radmacher, 2006. Direct measurement of the lamellipodial protrusive force in a migrating cell. *J Cell Biol* 174:767–772.
54. Kozlov, M. M., and A. D. Bershadsky, 2004. Processive capping by formin suggests a force-driven mechanism of actin polymerization. *J Cell Biol* 167:1011–1017.
55. Kubota, H., M. Miyazaki, T. Ogawa, T. Shimozaawa, K. Kinoshita Jr, and S. Ishiwata, 2017. Biphasic effect of profilin impacts the formin mediated force-sensing mechanism in actin polymerization. *Biophysical journal* 113:461–471.
56. Bryant, D., L. Clemens, and J. Allard, 2017. Computational simulation of formin-mediated actin polymerization predicts homologue-dependent mechanosensitivity. *Cytoskeleton* 74:29–39.
57. Courtemanche, N., J. Y. Lee, T. D. Pollard, and E. C. Greene, 2013. Tension modulates actin filament polymerization mediated by formin and profilin. *Proceedings of the National Academy of Sciences* 110:9752–9757.
58. Cao, L., M. Kerleau, E. L. Suzuki, H. Wioland, S. Jouet, B. Guichard, M. Lenz, G. Romet-Lemonne, and A. Jegou, 2018. Modulation of formin processivity by profilin and mechanical tension. *eLife* 7:e34176.
59. Kim, J. S., and S. X. Sun, 2009. Continuum modeling of forces in growing viscoelastic cytoskeletal networks. *Journal of theoretical biology* 256:596–606.
60. Mueller, J., G. Szep, M. Nemethova, I. de Vries, A. D. Lieber, C. Winkler, K. Kruse, J. V. Small, C. Schmeiser, K. Keren, et al., 2017. Load adaptation of lamellipodial actin networks. *Cell* 171:188–200.

APPENDIX

The effect of the filament-obstacle interaction on the force-velocity relation of a growing biopolymer

F. Motahari and A. E. Carlsson

Here we show results not including filament-tip fluctuations, and results incorporating slower diffusion.

Appendix A: Orthogonal Incidence without Filament-tip Fluctuations

When the filament grows orthogonal to the obstacle surface, the component of the filament-tip fluctuations along the direction of motion is greatly reduced. Therefore we ignore these fluctuations (see Fig A.1). Figure A.2 shows the resulting force-velocity relation for different pusher and puller potentials described in Figure 2. Notice that the stall force is smaller in this case because $\delta \cos(\theta)$ in Eq. 11 is replaced by δ . The BR relation remains an upper bound for the growth velocity, and the force-velocity relations for both the hard wall and soft wall are very similar to the BR relation. The $5k_B T$ well and spike potentials continue to have a zero-force velocity that is a substantial fraction of the free-filament velocity, with velocities decaying more rapidly than the BR prediction. In potentials with deep wells, the polymerization is slowed by roughly the same amount as in our baseline results (Figure 5). The tradeoff between polymerization rate and maximum sustainable pulling force is also preserved.

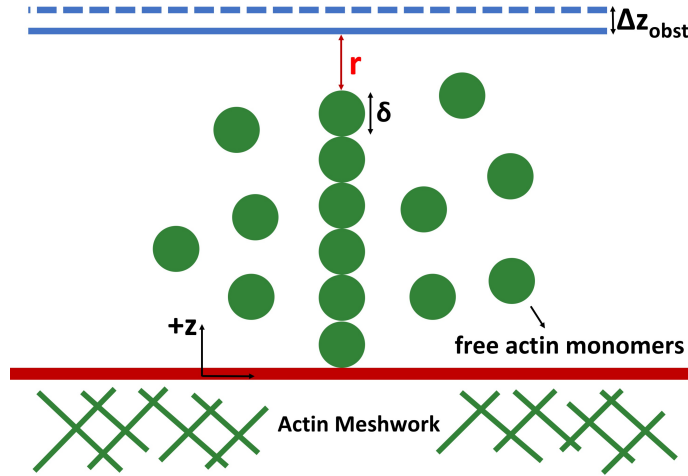


Figure A.1. Schematic of model of filament at orthogonal orientation. Filament tip fluctuations are ignored in this case.

Appendix B: Effect of Slow Obstacle Diffusion

We have repeated the force-velocity relations for the oblique filament case (45° orientation, including filament-tip fluctuations) with a smaller diffusion coefficient to see which findings in the main text depend strongly on the assumption of rapid diffusion. Figure B.1 shows results for a diffusion coefficient of $D = 21 \text{ nm}^2/\text{s}$. For this value of D , the dimensionless parameter characterizing diffusion $\delta^2 k_{on}/2D_{obst}$ has the value unity, so the effects of diffusion should be substantial. The polymerization rate for the more rapidly growing potentials - including the “Hard Wall”, the “Soft Wall”, and the “Puller” with $5 k_B T$ well depth, is slowed by about a factor of 2. As in the main text, the completely repulsive continuous potentials have force-velocity relations similar to the hard wall. However, the sharp drop in velocity for the $5k_B T$ potentials is eliminated, as expected from the analysis of Sec. 3.2 showing that this effect depends on rapid diffusion. The effect on the other force-velocity relations for deeper wells is smaller. The general shape of the force-velocity is unchanged, and there is no effect on the ordering of the curves.

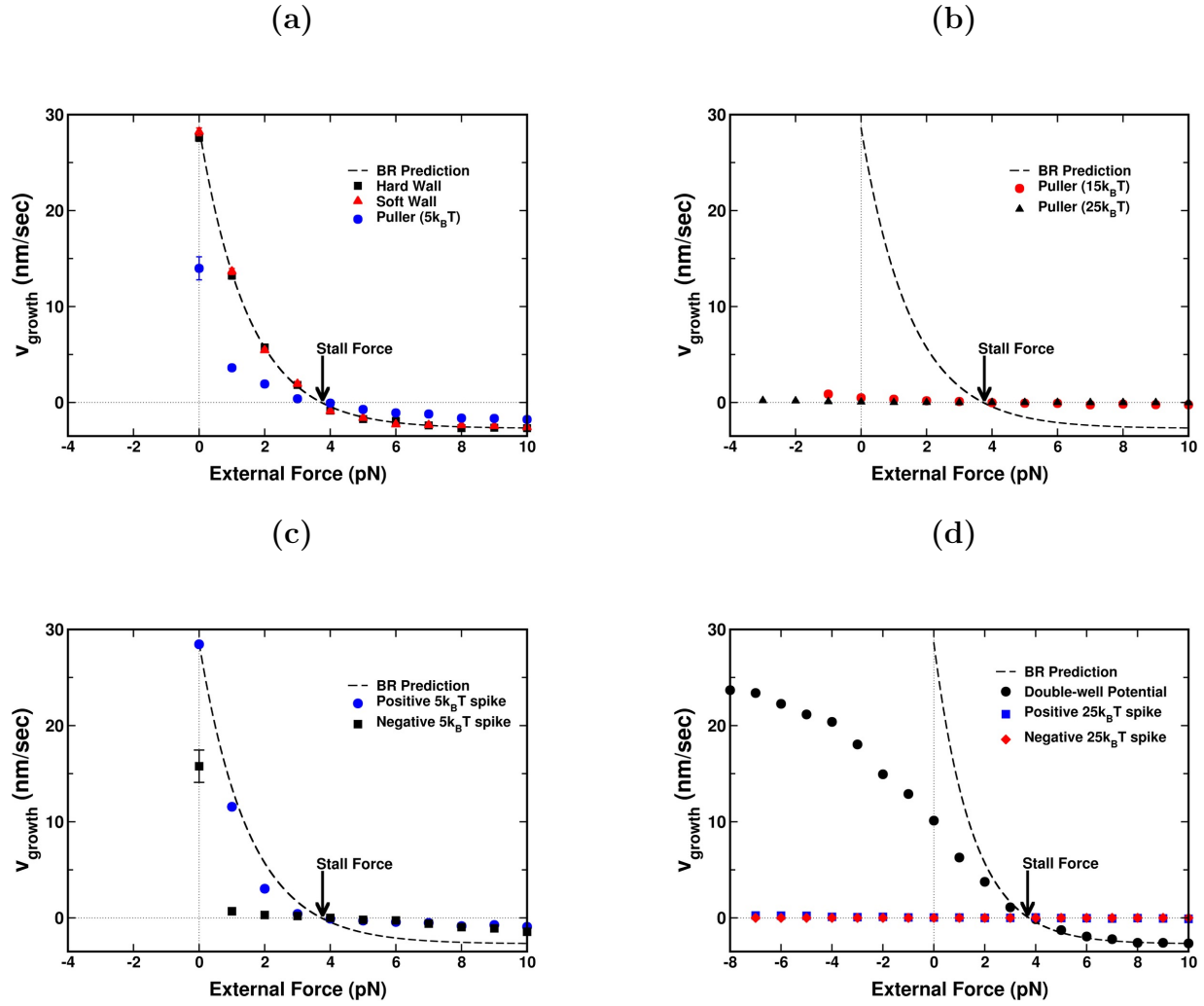


Figure A.2. Simulated force-velocity relation for several different forms of filament-obstacle interaction potential, for an orthogonal filament without tip fluctuations. Error bars are smaller than the symbols, except indicated; and they are calculated by finding the standard deviation of the mean for growth velocity, from 20 different simulations of 20 seconds. The dashed line is the prediction of the BR model at large diffusion constant values for obstacle. Data points for pullers are shown out to forces where they detach from the obstacle. Frames (a) and (b) are for potentials in Figure 2a, while (c) and (d) are for potentials in Figure 2b.

As Figure B.1 indicates, the effect of using a softer wall on the growth velocity is minimal even when diffusion is slow. For a still softer wall with decay coefficient $\kappa_1 = 0.2 \text{ nm}^{-1}$ (data not shown), the acceleration is about 10%.

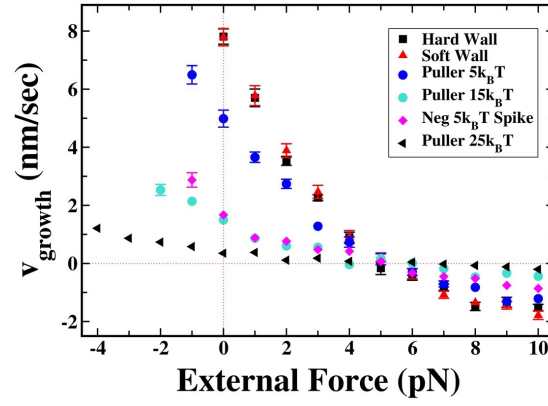


Figure B.1. Effect of reduced diffusion coefficient on force-velocity relation of growing actin filaments. Diffusion coefficient of $D = 21 \text{ nm}^2/\text{s}$. Force field parameters are as in Figure 5.